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(54) Title: METHOD OF PRODUCING ANTIHYPERCHOLESTEROLEMIC AGENTS

Lovastatin production genes

bavC:
Dehydrogenase OI
ORF5 | Esterase Ri tinger i ORF10: Metabolite transport NPKS

ORF13: ORF14: ORF15:
Cn Acetyl CoA Membrane
ORF12 finger II transport ORF15 transport

(57) Abstract

A method of increasing the production of lowestatin or monacolin J in a lowestatin-producing or non-lowestatin-producing organism is disclosed. In one embodiment, the method comprises the steps of transforming an organism with the A. terrest D4B segment, wherein the segment is translated and where an increase in lowestatin production occurs.

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METHOD OF PRODUCING ANTIHYPERCHOLESTEROLEMIC AGENTS

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CROSS-REFERENCES TO RELATED APPLICATION

Not applicable.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH AND DEVELOPMENT

This invention was made with United States government support awa;ded by the following agencies: NIH Grant No: AI43031. The United States has certain rights in this invention.

BACKGROUND OF THE INVENTION

- Cholesterol and other lipids are transported in body fluids by low-density lipoproteins (LDL) and high-density lipoproteins (HDL). Substances that effectuate mechanisms for lowering LDL-cholesterol may serve as effective antihypercholesterolemic agents because LDL
- 15 levels are positively correlated with the risk of coronary artery disease.
- MEVACOR (lovastatin; mevinolin) and ZOCOR (simvastatin) are members of a group of active antihypercholesterolemic agents that function by
- inhibiting the rate-limiting step in cellular cholesterol biosynthesis, namely the conversion of hydroxymethylglutarylcoenzyme A (HWG-CoA) into mevalonate by HMG-CoA reductase.

The general biosynthetic pathway of a naturally occurring HMG-CoA reductase inhibitor has been outlined by Moore, et al., who showed that the biosynthesis of

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mevinolin (lovastatin) by Aspergillus terreus ATCC 20542 begins with acetate and proceeds via a polyketide pathway (R.N. Moore, et al., J. Amer. Chem. Soc. 107:3694-3701, 1985). Endo, et al. described similar biosynthetic

5 pathways in Pencillium citrinum NRRL 8082 and Monascus ruber M-4681 (A.Y. Endo, et al., J. Antibiot. 38:444-448.

The recent commercial introduction of microbial HMG-CoA reductase inhibitors has fostered a need for high

yielding production processes. Methods of improving process yield have included scaling up the process, improving the culture medium and simplifying the isolation. Previous attempts to increase the biosynthesis of HMG-CoA reductase inhibitors at the level of gene expression have focused on increasing the concentration triol polyketide synthase (TPKS), a multifunctional protein with at least six activities as evidenced by the

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20 TPKS is believed to be the rate-limiting enzymatic activity(ies) in the biosynthesis of the HMG-CoA reductase inhibitor compounds.

product of the enzymatic activity (Moore, <u>supra</u>, 1985).

- U.S. patent 5,744,350 identifies a DNA encoding triol polyketide synthase (TPKS) from Aspergillus
- 25 terreus. "NPKS" is now preferred to TPKS as the acronym for "nonaketide polyketide synthase."

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SUMMARY OF THE INVENTION

In one embodiment, the present invention is a method of increasing the production of lovastatin in a lovastatin-producing organism. The method comprises the steps of transforming the organism with a nucleic acid sequence comprising the D4B segment, preferably comprising nucleotides 579 - 33,000 of SEQ ID NO:18 and 1 - 5,349 of SEQ ID NO:19. The nucleic acid sequence is transcribed and translated and an increase in lovastatin production occurs. Preferably, this increase is at least 2-fold.

In a preferred form of the present invention, the lovastatin-producing organism is selected from the group consisting A. terreus ATCC 20542 and ATCC 20541.

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In another embodiment, the method comprises the step of transforming the organism with the corresponding D4B segment isolated from a non-A. terreus lovastatin-producing organism.

In another embodiment, the present invention is a method of increasing the production of lovastatin in a lovastatin-producing organism, comprising the step of transforming the organism with the LovE gene, wherein the nucleic acid sequence is transcribed and translated and wherein an increase in lovastatin production occurs.

In another embodiment of the present invention, one may increase the production of monacolin J in a non-lovastatin-producing organism comprising the steps of

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transforming the organism with a nucleic acid sequence comprising the D4B segment. As a further step, one may additionally transform the organism with an entire LovF gene. If the entire LovF gene is added to the D4B segment, the organism will produce lovastatin.

In another embodiment, the present invention is the lovastatin production gene cluster, preferably SEQ ID NOs:18 and 19, and the individual genes comprising that cluster.

10 It is an object of the present invention to provide a method for increasing lovastatin and monacolin J production in both lovastatin-producing and non-lovastatin producing organisms.

Other objects, features and advantages of the present invention will become apparent after review of the specification, claims and drawings.

DESCRIPTION OF DRAWINGS

Fig. 1 is a diagram of lovastatin production genes
Fig. 2 is a schematic diagram of a hypothetical

20 mevinolin/lovastatin biosynthesis pathway.

Fig. 3 is a comparative diagram of statins.

Fig. 4 is a schematic drawing of plasmid pwHM1264/CB24A.

Fig. 5 is a schematic drawing of plasmid pWHM1424.

25 Fig. 6 is a schematic drawing of plasmid CD4B/pWHM1263.

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DESCRIPTION OF THE INVENTION

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The Examples below disclose the cloning and sequencing of a cluster of 17 genes from A. terreus ATCC 20542, a strain that natively produces lovastatin (See Fig. 1). These genes flank the NPKS gene, which is known to be required for lovastatin production (see, for

The DNA sequence of the cluster has been determined 10 and is disclosed below at SEQ ID NOs:18 and 19.

example, U.S. patent 5,744,350).

Mutations in four of the genes (P450I/LovA, SEQ ID NO:22; dehydrogenase/LovC, SEQ ID NO:24; esterase/LovD, SEQ ID NO:25; and SCPKS/LovF, SEQ ID NO:29) have been isolated and demonstrate that each of these four individual genes is required for lovastatin production. These genes are indicated with an X symbol in Fig. 1 and referred to herein as the "A. terreus lovastatin gene cluster."

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Another of the genes (Zn Finger I/LovE, SEQ ID NO:27) is thought to regulate the transcription of the contex genes and causes a notable increase in lovastatin production when reintroduced into A. terreus ATCC 20542.

Applicants have used the following convention in naming the genes and proteins of the present invention. The genes and proteins are first named with either an "ORF" or "Lov" prefix and then named either numerically or alphabetically. "Lov" signifies genes shown to be essential for lovastatin production. Applicants have

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also included a descriptor name that describes a probable function of the protein. For example, SEQ ID NO:1 is described as the "ORFI/esterase-like protein" because Applicants have compared the amino acid sequence to known esterases.

The portion of the gene cluster between

ORF1/esterase-like protein and the mid-region of LovF/SCPKS is referred to as the "D4B segment". The A.

terreus D4B segment is contained within a plasmid clone

deposited as ATCC 98876. As described below, other lovastatin-producing organisms contain an analogous D4B segment comprising analogous genes. The present

lovastatin-producing organisms. The arrangment of the genes within the D4B segment may be different in other

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invention comprises a "D4B segment" isolated from other

genes within the D4B segment may be different in other organisms. We predict that the genes within these other segments will have at least 80% homology, at the nucleic acid level, with the genes disclosed herein. We envision

that each of these lovastatin-producing organisms will 20 comprise within their genomes a LovA, LovB, LovC, LovD, LovE and LovF gene.

We have determined that the D4B segment will confer production of monocolin J if the genes are all expressed, as we show below in an example using A. nidulans. We

25 envision that adding the LovF gene to the D4B segment genes will result in the production of lovastatin in a non-lovastatin-producing organism.

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orientation, although, as Fig. 1 indicates, some of these cluster. SEQ ID NOs:18 and 19 are the entire DNA DNA sequences are in the inverted orientation in the cluster and include the introns. These DNA sequences are genomic DNA sequences of the various members of the gene sequence of the gene cluster. SEQ ID NOs:21-36 are the translation products of various members of the gene present invention. SEQ ID NOs:1-17 are predicted different protein and nucleic acid sequences of the actual cluster. reported in the Sequence Listing in the 5' - 3' Table 1, below, summarizes information regarding the

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TABLE 1

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| | | - Common | COMMENTS |
|----|--------------|---|--|
| | SEQ ID NO. | DESCRIPTION | COMMENIA |
| 15 | SEQ ID NO: I | Predicted amino acid sequence of ORF1/Esterase-like protein | Translation of 6 EXONS 6865- 6568, 6462-5884, 5320-4822, 4774- 3511, 3332-2372, 2301-1813 (reverse complement) FROM SEQ ID NO:18 |
| | SEQ ID NO: 2 | Predicted amino acid sequence of ORF2 | Translation of 1 EXON 7616-8602 FROM SEQ ID NO:18 |
| | SEQ ID NO: 3 | Predicted amino acid sequence of LovA/P4501 protein | Translation of 1 EXON 10951-9980 (reverse complement) FROM SEQ ID NO:18 |
| | SEQ ID NO: 4 | Predicted amino acid sequence of ORF5 | Translation of 1 EXON 22760- 21990 (reverse complement) FROM SEQ ID NO:18 |
| | SEQ ID NO: 5 | Predicted amino acid sequence of LovC/Dehydrogenase | Translation of 3 EXONS 23158- 23717, 23801-23912, 23991-24410 FROM SEQ ID NO:18 |
| 20 | SEQ ID NO: 6 | Predicted amino acid sequence of LovD/Esterase | Translation of 3 EXONS 26203- 26080, 26005-25017, 24938-24810 (reverse complement) FROM SEQ ID NO:18 |

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| | SE | SE | SE | SE | 5 SE | SE | SE | SE | SE | S. |
|--|--|--|--|---|--|--|--|--|--|-------------|
| SEQ ID NO: 16 | SEQ ID NO: 15 | SEQ ID NO: 14 | SEQ ID NO: 13 | SEQ ID NO: 12 | SEQ ID NO: 11 | SEQ ID NO: 10 | SEQ ID NO: 9 | SEQ ID NO: 8 | SEQ ID NO: 7 | SEQ ID NO. |
| Predicted amino acid sequence of ORF17/P450II protein | Predicted amino acid sequence of ORF 16/Membrane transport protein | Predicted amino acid sequence of ORF15 | Predicted amino acid sequence of ORF 14/Acetyl CoA transport protein | Predicted amino acid sequence of ORF13/Zn Finger II | Predicted amino acid sequence of ORF12 | Predicted amino acid sequence of LovF/ScPKS | Predicted amino acid sequence of ORF10/Metabolite transport | Predicted amino acid sequence of LovE/Zn Finger 1 | Predicted amino acid sequence of ORF8/HMG CoA Reductase | DESCRIPTION |
| Translation of 3 EXONS 28525- 27673, 27606-27284, 27211-26837 | Translation of 5 EXONS 24521- 24054, 23996-23936, 23876-23184, 23111-22977, 22924-22818 (reverse complement) FROM SEQ ID NO:19 | Translation of 2 EXONS 20332- 20574, 20631-21860 FROM SEQ ID NO:19 | Translation of 7 EXONS 19642- 19571, 19502-19427, 19352-19227, 19158-19011, 18956-18663, 18587- 18438, 18380-18341 (reverse complement) FROM SEQ ID NO:19 | Translation of 5 EXONS 16608- 16463, 16376-15572, 15519-15346, 15291-14825, 14767-14131 (reverse complement) FROM SEQ ID NO: 19 | Translation of 3 EXONS 13596- 13496, 13451-13063, 12968-12709 (reverse complement) FROM SEQ ID NO: 19 | Translation of 7 EXONS 4430- 4627, 4709-4795, 4870-4927, 4985- 5318, 5405-5912, 5986-6565, 6631- 12464 FROM SEQ ID NO:19 | Translation of 8 EXONS 1400- 1452, 1619-1695. 1770-1996. 2063- 2088, 2154-2225. 2332-2865. 2939- 3099, 3180-3560 FROM SEQ ID NO:19 | Translation of 1 EXON 31360- 32871 FROM SEQ ID NO:18 | Translation of 5 EXONS 30062. 29882, 29803-29745, 29664-27119. 27035-26779, 26722-26559 (reverse complement) FROM SEQ ID NO:18 | COMMENTS |

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| COMMENTS | Translation of 2 EXONS 29826- 30995, 31054-31328 (incomplete) FROM SEQ ID NO:19 | | | Start = 6865 Stop = 1813 SEQ ID NO:18 | Start = 7616 Stop = 8602 SEQ ID NO:18 | Start = 10951 Stop = 9980 SEQ ID NO:18 | Start = 22760 Stop = 21990 SEQ ID NO:18 | Start = 23158 Stop = 24410 SEQ ID NO:18 | Start = 24810 Stop = 26203 SEQ ID NO:18 | Start = 30062 Stop = 26559 SEQ ID NO:18 | Start = 31360 Stop = 32871 SEQ ID NO:18 | Start = 1400 Stop = 3560 SEQ ID NO:19 | Start = 4430 Stop = 12464 SEQ ID NO:19 | Start = 13596 Stop = 12709 SEQ ID NO:19 |
|-------------|---|---|--|---|---|--|---|---|---|---|---|---|--|---|
| DESCRIPTION | Predicted amino acid sequence of ORF18 (incomplete) | DNA sequence of gene cluster- first 33,000 nucleotides | DNA sequence of cluster- nucleotides 33,001-64,328 (renumbered 1-31,328) | DNA sequence of ORF1/Esterase-like gene | DNA sequence of ORF2 | DNA sequence of LovA/P450I gene | DNA sequence of ORF5 | DNA sequence of LovC/Dehydrogenese | DNA sequence of LovD/Esterase | DNA sequence of ORF8/HMG CoA Reductase | DNA sequence of LovE/Zn Finger I | DNA sequence of ORF10/Metabolite transport | DNA sequence of LovF/ScPKS | DNA sequence of ORF12 |
| SEQ ID NO. | SEQ ID NO: 17 | SEQ ID NO: 18 | SEQ ID NO: 19 | SEQ ID NO: 20 | SEQ ID NO: 21 | SEQ ID NO: 22 | SEQ ID NO: 23 | SEQ ID NO: 24 | SEQ ID NO: 25 | SEQ ID NO: 26 | SEQ ID NO: 27 | SEQ ID NO: 28 | SEQ ID NO: 29 | SEQ ID NO: 30 |

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| | SEQ ID NO. | DESCRIPTION | COMMENTS |
|-----|---------------|---|--|
| *** | SEQ ID NO: 31 | DNA sequence of ORF13/Zn Finger II | Start = 16608 Stop = 14131 SEQ ID NO:19 |
| | SEQ ID NO: 32 | DNA sequence of ORF14/Acetyl CoA transport gene | Start = 19642 Stop = 18341 SEQ ID NO:19 |
| | SEQ ID NO: 33 | DNA sequence of ORF15 | Start = 20332 Stop = 21860 SEQ ID NO:19 |
| | SEQ ID NO: 34 | DNA sequence of ORF16/Membrane transport protein | Start = 24521 Stop = 22818 SEQ ID NO:19 |
| | SEQ ID NO: 35 | DNA sequence of ORF17/P450II gene | Start = 28525 Stop = 26837 SEQ ID NO:19 |
| | SEQ ID NO: 36 | DNA sequence of ORF18 (incomplete) | Start = 29826 to 31328 (incomplete) SEQ ID NO:19 |

Table 1 also notes the translation start and stop points in the various gene sequences.

10 The sequence of the NPKS gene is not listed in SEQ ID NOs:21-36. This gene is characterized in U.S. patent 5,744,350. However, SEQ ID NOs:18 and 19 do contain the sequence of the NPKS gene within the context of the entire gene cluster.

15 To perform many embodiments of the present invention, one will need to recreate various genes or a portion of the gene cluster described herein. Applicants have provided sequence data in the Sequence Listing sufficient to allow one of skill in the art to construct numerous probes suitable to recreate the genes from an A.

20 numerous probes suitable to recreate the genes from an A.

cerreus genomic library. Applicants have also described below various methods for isolating A. terreus DNA.

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Additionally, Applicants have deposited ATCC

Accession No. ATCC 98876, which contains clone pWHM1263 (cD4B) and ATCC Accession No. ATCC 98877 which contains clone pWHM1265 (CB2A4). Both plasmids are described in more detail below. Fig. 4 describes clone

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CB2A4/pWHM1265, and Fig. 6 describes clone CB4B/pWHM1263.

Fig. 1 also indicates the boundaries of the D4B and B2A4 clones.

The clones and their inserts may be prepared from

the ATCC deposits by methods known to those of skill in the art. The DNA from the clones may be isolated and any gene within the gene cluster may be isolated and utilized.

Increasing the Production of Lovastatin by Lovastatinproducing Fungi and Yeast

In one embodiment, the present invention is a method of increasing the production of lovastatin in a lovastatin-producing fungi and yeast, preferably A. terreus ATCC20542 and ATCC20541. Other examples of

20 suitable lovastatin-producing fungi and yeast are listed in Table 2, below.

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| | Microorganisms other than A. terreus reported to produce lovastatin (mavinglin) |
|----|--|
| | Monascus (17 of 124 strains screened) specias' |
| ທ | M. ruber M. purpureus M. pilosus M. vitreus M. pubigerus |
| 10 | Penicilium sp.¹ Hypamyces sp. Doratomyces sp. Phona sp. Eupenicilium sp. |
| 15 | Cuperinamen ap. Gymnoascus sp. Trichoderma sp. |
| | Pichia labacensis² Candida cariosilognicola |
| 20 | Aspergilus oryzee ³ Doratomyces stemonitis Paecilomyces virioti Penicilium citronum Penicilium citronum Penicilium citronum Penicilium citronum |
| | The state of the s |
| 30 | P. Juriova I. Marinkova, V. Kren. Secondary Metabolites of the fungus Monascus: a review. J. Ind. Microbiol. 16:163-170 and references cited therein (1996). N. Gunde-Cimerman, A. Phemeriusa and A. Cimerman. A hydroxymethylglutaryl-CoA reductase inhibitor synthesized by yeasts. EEMS Microbiol. Lett. 132:39-43 (1995). A.A. Shindia. Mevinoin production by some tungl. Folio Microbiol. 42:477-480 (1997). |

By "increasing the production" we mean that the amount of lovastatin produced is increased by at least 2-fold, preferably by at least 5-fold. The examples below demonstrate two preferred methods for analyzing strains for lovastatin production. In method A, the spore suspension is inoculated into a flask of SEED medium and grown. The resulting seed culture is used to inoculate FM media and grown for six days. In fermentation method

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B, one inoculates 50 ml of RPM media and grows this larger culture for 7 days.

Both cultures are extracted, pH adjusted, mixed with ethyl acetate and shaken for two hours. For analysis, 1 ml of the ethyl acetate layer is dried under a nitrogen stream and resuspended in methanol. For TLC analysis, a small amount of the extract is run on C18 reverse phase TLC plates in a solvent system of methanol; 0.1% phosphoric acid. The TLC plates are developed by spraying with phosphomolybdic acid in methanol and heating with a heat gun. The extracts are compared with

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If one wishes HPLC analysis, the examples below describe the use of a Waters Nova-Pak C18 column used with a solvent system of acetonitrile and phosphoric acid. A Waters 996 Photodiode Array Detector will detect the metabolites. Lovastatin was detected at 238 nm.

authentic lovastatin, monacolin J, monacolin L and

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dihydromonocolon L.

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In one embodiment, one would transform a lovastatin20 producing fungi or yeast with the lovE/zinc finger I
gene, preferably comprising the nucleotides of SEQ ID
NO:27. The examples below predict that this will result
in an increase of at least 5-7 fold. Preferably, the
increase will be at least 2.0-fold.

One may also transform a lovastatin-producing fungi or yeast with the LovE gene isolated from other lovastatin-producing fungi or yeast. One may obtain this

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gene by use of a probe derived from SEQ ID NO:27 by methods known to those of skill in the art.

One may also transform lovastatin-producing fungiand yeast with the D4B segment of the lovastatin production gene cluster (see Fig. 1), preferably as found in ATC accession number 98876. Alternatively, one may transform lovastatin-producing fungi or yeast with the entire gene cluster, as diagramed in Fig. 1.

We envision that to successfully increase lovastatin production, one may also wish to transform less than the entire gene cluster. Preferably, one may determine what the smallest possible segment is by deleting various portions of the gene cluster and determining whether lovastatin production is continually increased.

15 Similarly, if one begins with the D4B segment, one may delete various portions for the segment and determine whether lovastatin production is continually increased by at least 2-fold.

Modification of the LovB/NPKS gene would produce other HMG CoA inhibitors. For example, Fig. 3 diagrams the relationship between mevastatin, lovastatin, simvastatin and pravastatin. In one example, the methyl transferase domain of the NPKS gene may be replaced with an inactive form to make pravastatin. The HMG-CoA

reductase inhibitors within this invention include, but are not limited to, compactin (ML-236B), lovastatin, simvastatin, pravastatin and mevastatin.

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In another embodiment of the present invention, one may transform a lovastatin-producing organism with the genes described above and obtain the production of an HMG COA reductase inhibitor with a structure different from monacolin J, monacolin L or lovastatin. Alterations in the side chain attached to C8 are the most likely possibility but other alterations may occur. These alterations would happen through the native biochemistry of the organism.

10 If one wishes to express the A. terreus genes in yeast, one may wish to consult examples in which others have engineered fungal secondary metabolism genes for expression in yeast. (See for example, J. T. Kealey, £L al., Proc. Natl. Acad. Sci. USA 95:505-509 (1998)). The exact approach could be used with the NPKS (LovB) and ScpKS (LovF) genes, and a somewhat simpler approach with the other lovastatin genes in their cDNA forms.

Production of HMG-CoA Reductase Inhibitors by Fungi and Yeast That Do Not Natively Produce Inhibitors

- In another embodiment, the present invention is the production of HMG-CoA reductase inhibitors, such as lovastatin, by fungi and yeast that do not natively produce lovastatin. An example of a suitable fungi or yeast is A. nidulans and S. cerevisiae, respectively.
- 25 For this embodiment one preferably transforms the genes within the D4B segment into the non-inhibitor-producing strain. By this method, one would produce

monacolin J (See Fig. 2) which could be chemically

converted to lovastatin by one of skill in the art.

Monacolin J, in its lactone form obtained by treatment with anhydrous acid under dehydrative conditions, is preferably treated with a derivative of (2S)-2-methybutyric acid, in which the carboxyl group has

- been suitable activated for undergoing esterification, and the resulting lovastatin is isolated by conventional methods. For example, see WO 33538, U.S. patent
- 10 4,444,784 and <u>J. Med. Chem.</u> 29:849 (1986). These are citations for synthesis of simvastatin from monacolin J.

 One would use the same method, but use the (2S)-2methylbutyrate derivative to make lovastatin.

In another embodiment of the present invention, one would transform the genes within the D4B segment, including an entire LovF/SCPKS gene, into the non-inhibitor-producing organism. By this method, one would produce lovastatin in a non-lovastatin-producing organism.

In another embodiment of the present invention, one may transform a non-lovastatin-producing organism with the genes described above and obtain the production of an HMG CoA reductase inhibitor with a structure different from monacolin J, monacolin L or lovastatin, as described above.

Modification of the LovB/NPKS gene would produce other inhibitors. For example, Fig. 3 diagrams the relationship between mevastatin, lovastatin, simvastatin

and pravastatin. In one example, the methyl transferase domain of the NPKS gene may be replaced with an inactive form to make pravastatin. The HMG-CoA reductase inhibitors within this invention include, but are not

5 limited to, compactin (ML-236B), lovastatin, simvastatin, pravastatin and mevastatin.

Production of Intermediate Materials

In another embodiment, the present invention is a method of isolating intermediate materials in the production of lovastatin and analogs such as mevastatin

and simvastatin. For example, the Examples below demonstrate the disruption of the lovastatin projection gene cluster with mutagenized LovC, LovD, LovF, LovA or LovB genes. Disruption of many of these genetic elements

accumulation of intermediate materials. Therefore, to practice this embodiment of the present invention, one would transform a suitable lovastatin-producing host with a mutagenized gene within the D4B segment, as described

:0 below. Many other mutations would be suitable to destroy the function of LovC, LovD, LovF, LovA or LovB. All that

is necessary is these genes be disrupted to the extent

25 Production of Lovastatin Analogs

that they are non-functional.

In another embodiment, the present invention provides a method for engineering the production of

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lovastatin analogs in such organisms as fungi or yeast, using monacolin J as the starting point.

Isolated DNA Segments

In another embodiment, the present invention is a DNA segment capable of conferring lovastatin or monacolin J production or increase in lovastatin or monacolin J production in yeast or fungi. In a preferred example, this segment is the "D4B segment" that is deposited at ATCC 98876. The nucleotide sequence of this segment is found in residues 579 - 33,000 of SEQ ID NO:18 and residues 1 - 5,349 of SEQ ID NO:19.

In another embodiment, the present invention is the entire A. terreus lovastatin gene cluster, as exemplified by SEQ ID NOs:18 and 19 and ATCC deposits 98876 and 98877.

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The present invention is also the individual genes that make up the A. terreus lovastatin gene cluster.

Therefore, the present invention is a nucleic acid segment selected from the group of consisting of SEQ ID NOS:20 - 36. Preferably, the present invention is the

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- NOS:20 36. Preferably, the present invention is the coding region found within SEQ ID NOS:20 36 and described in Table 1. The present invention is also a mutagenized version of SEQ ID NOS:22, 24, 25 and 29, wherein the gene is mutagenized to be non-functional in
 - 25 terms of lovastatin or monacolin J production.

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Organisms with Increased Loyastatin or Monacolin J Production

In another embodiment, the present invention are the organisms described above. These organisms include lovastatin-producing organisms, preferably yeast and fungi, that have been engineered to display at least a 2-fold increase in lovastatin or monacolin J production. The organisms also include non-lovastatin-producing organisms, preferably yeast or fungi, that have been

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Antifungal Compounds

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engineered to produce monacolin J or lovastatin.

Applicants note that lovastatin, monocolin J, monocolin L and dihydromonocolin L all have varying degrees of antifungal activity. Applicants envision that the present invention is also useful for providing antifungal compounds and organisms engineered to express antifungal compounds. Preferably, one would measure the antifungal properties of a compound in the manner of N. Lomovskaya, et al., Microbiology 143:875-883, 1997.

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R. Ikeura, et al., J. Antibiotics 41:1148, 1988. The

same general methods could be used for all fungi. Both

Measurement of inhibition of yeast growth can be found in

of these references are hereby incorporated by reference

EXAMPLES

. General Methods and Procedures

Construction of an A. terreus ATCC20542 genomic library.
A. terreus ATCC20542 genomic DNA was partially

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- digested with Sau3AI so as to produce an average fragment size of 40 50 kb. The partially digested genomic DNA was then separated on a sucrose gradient and the 40 50 kb fraction was collected. Cosmid AN26 (Taylor and Borgmann, Fungal Genet. Newsletter 43, 1996) was prepared by digestion with ClaI, dephosphorylated with CIP, then
- digested with BamHI to create the two cosmid arms.

 Ligation reactions with genomic DNA fragments and cosmid arms were optimized and packaged using Gigapack III XL packaging extract (Stratagene). The packaged cosmid library was infected into E. coli JM109 and plated out onto LB agar (Sambrook, gt al., Molecular Cloning. A Laboratory Manual. 2nd ed. Cold Spring Harbour Laboratory Press, 1989; other standard methods used can be found here also) with ampicillin (50 µg/ml) plates.
- 20 After checking for the presence of insert DNA in a selection of clones, 5000 colonies were picked into LB plus 50 μ g/ml ampicillin filled microtitre plates and grown overnight at 37°C. The colonies were replica plated onto nylon membranes (Amersham Hybond-N).
- 25 Glycerol was added at a final concentration of 15% (v/v) to the microtitre plates and these were stored at -70°C.

Isolation of genomic clones containing the lovastatin biosynthesis cluster.

A 2.8 kb EcoRI fragment from pTPKS100 containing part of the NPKS gene (Vinci, et al., U.S. Patent No.

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- 5,744,350) was gel-isolated and labelled with digoxigenin labelled fragment was hybridized (65°C, 5x SSC) with the library, then washed (65°C, 0.1x SSC). Two positive nylon membranes containing the A. terreus genomic using the Genius Kit II (Boehringer Mannheim).
- Soulevard, Menassas, VA 20110) at accession number ATCC clones were identified, PWHM1263 (cD4B) and pWHM1264 (American Type Culture Collection, 10801 University (cJ3A). Two of these clones, pWHM1263 (cD4B) and WHM1265 (cB2A4), have been deposited in the ATCC 10
- conditions of the Budapest Treaty. The presence of the 98876 and 98877, respectively, under the terms and NPKS gene was confirmed initially by restriction digestion and later by DNA sequencing. 15

PWHM1271 (CQ1F1) from upstream of NPKS. All these clones hybridization process using labelled fragments from both isolation of pWHM1265-1270 (cB2A4, cL3E2, cJ3B5, cO2B5, were transformed into E. coli strain STBL2 (Stratagene) ends of the insert in pWHM1263. This resulted in the Overlapping clones were found by repeating the 2R3B2, cW3B1) from downstream of the NPKS gene and to help prevent rearrangements. 20 25

This clone contains an insert of approximately 43 kb in Fig. 4 is a diagram of the cB2A4/pWHM1265 clone.

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SEQ ID NO:19 and 10 - 14 kb of uncharacterized DNA. Fig. nucleotides 4988 of SEQ ID NO:19 to nucleotide 31,328 of nucleotides 579 - 33,000 of SEQ ID NO:18 and nucleotides AN26 and includes the nucleotide sequence from at least 6 is a schematic diagram of cD4B/pWHM1263. This clone contains a 37,770 bp insert in AN26 and contains 1 - 5,349 of SEQ ID NO:19.

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Sequencing strategy and analysis.

A series of overlapping subclones (pWHM1272-

- Cycle sequencing was carried out using the AmpliTaq FS or BigDye reagents (ABI) and were analyzed using a ABI model prepared using the QiaPrep spin miniprep kit (Qiagen). pWHM1415) were constructed in pSPORT1 (Gibco-BRL) and pGEM3 (Promega). Plasmid DNAs for sequencing were 10
- by synthesis of 18-22 bp oligonucleotide primers based on the sequenced DNA strand, with the help of the Oligo 4.05 Primer walking was performed DNA was sequenced at least once on both strands. Direct program (National Biosciences, Inc.). Every region of 373 or 377 DNA Sequencer. 15
- existed. DNA sequence analysis and manipulations were confirm adjoining regions where no overlapping clones sequencing of cosmids and PCR products was used to software. Assignments of putative ORFS, including performed using SegMan (DNASTAR) and SegEd (ABI)

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2.0 searches (Atschul, et al., Nucl., Acida Res. 25:3389putative introns, were performed with the aid of BLAST programs (Program Manual for the Wisconsin Package, 3402, 1997), and the Genetics Computer Group (GCG)

WO 00/37629 Version 8, September 1994, Genetics Computer Group, PCT/US99/29583

Madison, WI), version 8.1.

ORF3) mutants. Isolation and characterization of lovF (ScPKS, ORF11), lovD (EST1, ORF7), lovC (DH, ORF6), and lovA (P4501,

Acc65I - HindIII fragment, into pPLOA (Vinci, et al., fragment was then subcloned from this vector, as an from pWHM1265 into pSPORT1 to give pWHM1291. The ScPKS EcoRI fragment internal to the lovF gene was subcloned To disrupt the polyketide synthase gene, lovF, a 1.7

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U.S. Parent No. 5,744,350) to give pWHM1416. This vector InVitrogen) resistance gene for selection in A. terreus. contains the phleomycin (Zeocin, obtained from

15 A. terreus ATCC20542 was then transformed to Zeocin resistance with this plasmid as described below WMH1731, lovastatin production was abolished and a new described below (Method A). In one of the transformants Transformants were screened for lovastatin production as

20 25 partially purify the major product which was then monacolin J on TLC and HPLC according to the methods compound accumulated. This new compound comigrated with pattern as authentic monacolin J was observed. To described below. Semi-preparative HPLC was used to analyzed by HPLC - MS. The same mass and fragmentation

5 WO 00/37629 with BamHI and HindIII, electrophoresed on an agarose gel bands at 6.5 kb and 2.2 kb for BamHI and 11 kb and 7.8 kb at 4.2 kb for BamHI and 11.5 kb for HindIII. previously. The wild-type strain had hybridizing bands membrane was hybridized with the 1.7 kb EcoRI fragment single copy of pWHM1416 at the lovF locus. for HindIII confirming the homologous integration of a predicted, the WMH1731 mutant strain had hybridizing (Boehringer Mannheim) using the conditions described from pWHM1416 labelled using the Genius II kit and capillary blotted onto a nylon membrane. The

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lovD

15 25 20 digested with HindIII and BamHI and the 6.6 kb fragment PWHM1263 was subcloned into pSPORT1 to give PWHM1274. and the 2.1 kb fragment containing the phleomycin 1.8 kb fragment was isolated. The plasmid was also like gene, lovD, a 4.8 kb NotI - EcoRI fragment from and the end of the lovD gene was isolated. This plasmid was isolated. This plasmid was digested with HindIII and BsiWI and a were ligated together and used to transform competent E. resistance marker was purified. These three fragments was linearized by digestion with XbaI or RsrII before coli cells. The expected plasmid, pWHM1417, containing phleomycin resistance gene flanked by the beginning To disrupt the putative esterase/carboxypeptidasepPLOA was digested with BamHI and Acc65I

the WMH1731 mutant strain.

The genomic DNA was digested

DNA was prepared from wild-type A. terreus ATCC20542 and confirm the disruption of the lovF gene, total genomic

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In one of the Transformants were screened for lovastatin being used to transform A. terreus ATCC20542 to Zeocin transformants, WMH1732, lovastatin production was production as described below (Method A). resistance.

- preparative HPLC was used to partially purify the major mass and fragmentation pattern as authentic monacolin J The same gene, total genomic DNA was prepared from wild type A. terreus ATCC20542 and the WMH1732 mutant strain. The compound comigrated with monacolin J on TLC and HPLC was observed. To confirm the disruption of the lovD abolished and a new compound accumulated. This new according to the methods described below. Semiproduct which was then analyzed by HPLC - MS. 10
 - As predicted the mutant strain had hybridizing bands at 9 genomic DNA was digested with ApaI, run out on an agarose using the conditions described previously. The wild-type strain had hybridizing bands at 9 kb, 8.4 kb and 1.5 kb. fragment from pWHM1274 labelled using the Genius II kit membrane was hybridized with the 4.8 kb NotI - EcoRI integration of a single copy of pWHM1417 at the lovD kb, 8 kb, 3 kb and 1.5 kb confirming the homologous gel and capillary blotted onto a nylon membrane. 15 20
- lova 25

into pGEM3 to give pWHM1272. From this plasmid a 2.1 kb To disrupt the cytochrome P450 I gene, lovA, an 11 kb Acc651 - EcoRI fragment from pWHM1263 was subcloned

lovastatin production was abolished and two new compounds Apal - SnaBl fragment was purified and ligated to Apal . out on an agarose gel, and capillary blotted onto a nylon membrane. The membrane was hybridized with the 6 kb ApaI EcoRV digested pPLOA to give p450Phleo (pWHM1418). From wild-type strain had hybridizing bands at 2.0 kb, 1.9 kb this plasmid a 4.2 kb ApaI - NotI fragment was purified accumulated. Genomic DNA was prepared from this strain - Kpnl fragment from pWHM1419 labelled using the Genius and from A. terreus ATCC20542, digested with EagI, run screened for lovastatin production as described below plasmid was then digested with KpnI and ApaI and the and ligated with a 1.8 kb Eagl - KpnI fragment from resulting fragment was used to transform A. terreus ATCC20542 to Zeocin resistance. Transformants were p450Dphleo (pWHM1419) which contains the lovA gene (Method A). In one of the transformants, WMH1733, II kit using the conditions described previously. PWHM1272 and ApaI - KpnI digested pGEM7 to give disrupted by the phleomycin resistance gene. 10 15 20

-26-

and 1.1 kb. Mutant strain WMH1733 had hybridizing bands

nomologous integration of a single copy of the fragment

from pWHM1419 at the lovA locus.

at 2.5 kb, 2.0 kb, 1.1 kb and 0.7 kb confirming the

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LOVC

To disrupt the dehydrogenase-like gene, lovC, a 2 kb EcoRI - BglII fragment from pTPKS100 was ligated with a 1.7 kb EcoRI - SacI fragment from pWHM1274 and BglII -

- 5 SacI digested litmus 28 (New England Biolabs) to produce pDH1 (pWHM1420). Another plasmid pDH2 (pWHM1421) was constructed from a 2.2 kb Acc65I SacI fragment from pWHM1274, a 2.1 kb HindIII SacI fragment from pPLOA containing the phleomycin resistance gene and HindIII -
- 10 Acc65I digested litmus 28. The disruption vector pDH-dis
 (pWHM1422) was constructed by ligating together a 2.5 kb

 BglII HpaI fragment from pWHM1420, a 4.3 kb EcoRV
 KpnI fragment from pWHM1421 and BglII KpnI digested

 litmus 28. This plasmid was digested with BglII and KpnI

 15 and the resulting 6.8 kb fragment was used to transform

 A. terreus ATCC20542 to Zeocin resistance. Transformants

 were screened for lovastatin production as described
- 20 Genomic DNA was prepared from these strains and from A. terreus ATCC20542, digested with EagI, run out on an agarose gel, and capillary blotted onto a nylon membrane. The membrane was hybridized with the 6.8 kb Bgl II- KpnI fragment from pWHM1422 labelled using the Genius II kit

below (Method A). In two of the transformants, WMH1734

and WMH1735, lovastatin production was abolished.

25 using the conditions described previously. The wild type strain had hybridizing bands at 5 kb, 1.5 kb and 1.3 kb.

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Mutant strain WMH1734 had hybridizing bands at 4.9 kb,

1.3 kb, 1.0 kb and 0.7 kb confirming the homologous integration of a single copy of the fragment from pWHM1422 at the lovC locus. The other mutant strain,

5 WMH1735, had a similar banding pattern but with additional hybridizing bands indicating that multiple integration events had occurred, one of which was at the lovC locus.

Construction and characterization of the A. terreus 10 strain with extra copies of lovE.

A 10.4 kb Not1- EcoRI fragment containing the putative regulatory gene, lovE was subcloned from pwHM1263 to pSPORT1 to give pwHM1276. From this plasmid a 3.9 kb HindIII - BamHI fragment was subcloned into

- 15 pGEM7 to give pWHM1423. The regulatory gene was subcloned from this vector into pPLOA as an SstI XbaRI fragment to give pWHM1424 (Fig. 5). pWHM1424 contains nucleotides 30,055 33,000 from SEQ ID NO:18 and nucleotides 1 1,026 from SEQ ID NO:19.
- into A. terreus ATCC20542 by transformation to Zeocin resistance with pWHM1424. Transformants were fermented (method A) and screened for lovastatin production initially by TLC analysis. Most of the transformants appeared to be producing significantly more lovastatin from than the wild-type strain. The yields of lovastatin from the two transformant strains, WMH1736 and WMH1737, which had the most elevated levels compared to the wild-type

was quantified by HPLC as described below. These were WO 00/37629

found to produce 7-fold and 5-fold more lovastatin than the A. terreus ATCC20542 strain.

Because of the way that the DNA integrates

Ď genotypically and phenotypically. However, some will (ectopically), each transformant is or can be unique, overproducers; others may exhibit no difference, for unknown reasons.

Heterologous expression of the lovastatin biosynthesis genes. 20

frame was ligated into pAL3 (Waring, <u>et al</u>., Gene 79:119, fragment from pTPKS100 containing the NPKS open reading To place the NPKS gene (lovB) under the control of the inducible alcA promoter, the 11.5 kb KpnI - AvrII

- resulting plasmid was designated pAL3TPKS (WHM1425). The upstream of the translational start codon and a Agel site polymerase chain reaction was used to amplify the NPKS gene sequence between the NPKS promoter region just 1989) previously digested with KpnI and XbaI. 12
- from the A. terreus system. Amplification was performed start codon allowing the NPKS to be placed against the introduced a KpnI site 31 bases from the translational alcA promoter but also incorporating upstream elements internal to NPKS. The design of the forward primer 20
- using Vent DNA polymerase with pTPKS100 as template and 1 μ mol of each primer in a final volume of 100 μ l using the manufacturer's buffer recommendations. After an initial 25

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was achieved with 30 cycles of 95°C for 1 minute; 55°C for The final cycle was denaturation cycle of 10 minutes at 95°C amplification followed by 10 minutes at 72°C to ensure complete 1 minute and 72°C for 1.5 minutes.

- pAL3TPKSNT (pWHM1426). The region introduced by PCR was transform A. nidulans strain A722 (Fungal Genetics Stock digested with KpnI and AgeI and ligated into pWHM1425 that had been digested with the same enzymes and gel sequenced on a ABI automated DNA sequencer to ensure polymerization. The amplified product (1.7 kb) was sequence fidelity. This plasmid was then used to isolated. The resulting plasmid was designated 9
- spore suspension (10° c.f.u./ml) in 50 ml YEPD in a 250 ml This was then grown for 20 hours at 250 Transformants were grown by inoculating 0.5 ml of Incubator Shaker). The mycelia were then harvested rpm and 37°C (New Brunswick Scientific Series 25 unbaffled flask. 15

Centre) to uridine prototrophy.

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filtration through Miracloth (Calbiochem), rinsed with

- filtration using Miracloth (Calbiochem), squeezed as dry mM cyclopentanone and grown for a further 20 hours under ml unbaffled flasks containing 50 ml AMM + lactose + 10 sterile, distilled water, and inoculated into fresh 250 the same conditions. The mycelia were harvested by 20
 - extracts for SDS-PAGE and western analysis were prepared as described in Kennedy and Turner, Molec. Gen. Genet. as possible and frozen in liquid nitrogen. Protein (1996), 253:189-197, 1996. 25

WO 00/37629 protein (>200 kDa) visible on a SDS-PAGE gel that cross reacted with the affinity purified NPKS antibodies (Panlabs). This strain WMH1738 was transformed to One transformant, WMH1738, was shown to have a large PCT/US99/29583

- hygromycin B resistance with pWHM1263. Transformant analysis. Both of these strains were found to be two strains WMH1739 and WMH1740 were chosen for further the production of new metabolites as described below and colonies were screened for lovastatin resistance and for
- 10 significantly resistant (up to 100 μ g/ml on solid media) 5, 10, 50 and 100 $\mu g/ml$ and incubating at 37°C. Strains solid AMM plates containing lovastatin at 0, 0.1, 0.5, 1 analyzed by streaking 10 μl of a spore suspension on to lovastatin compared with the host strain. This was
- 15 20 WMH1739 and WMH1740 were compared to strains WMH1741 and lovastatin concentrations whereas strains WMH1741 and hygromycin resistance with AN26. Strains WMH1739 and -1740 exhibited no inhibition of growth at any of these WMH1742 which were derivatives of WMH1738 transformed to
- 1742 showed slight inhibition of grown at 5 $\mu g/ml$ and lovastatin-producing conditions using fermentation method almost complete growth inhibition at 50 $\mu \mathrm{g/ml}$. The two B and extracts were analyzed for lovastatin related lovastatin resistant strains were fermented in
- 25 analysis by the methods described below. Semimetabolites as described below. Both strains were found to both comigrated with monacolin J on TLC and HPLC to produce new metabolites. One compound that was common

WO 00/37629 preparative HPLC was used to partially purify some of the strains, comigrated with monacolin L on TLC and HPLC this compound, which was then analyzed by HPLC - MS. It monacolin J. The other compound, found in only one of had the same mass and fragmentation pattern as authentic PCT/US99/29583

Solid medium for growth of A. terreus

For the generation of spore suspensions A. terreus strains were grown on CM agar at 30°C for 4 to 5 days.

10 3 Agar (for CM liquid medium the agar was omitted): 50 ml Clutterbuck's salts (Vinci, et al., U.S.

20 15 Patent No. 5,744,350)

2 ml Vogel's trace elements (Vinci, <u>et al</u>., U.S. Patent No. 5,744,350)

0.5% Difco Bacto tryptone
0.5% Difco Bacto yeast extract 1% glucose
2% Difco Bacto agar
in 1 liter of distilled water

Clutterbuck's salts:

12% NANO, 1.02% KCl 1.04% MgSO,.7H₂O 3.04% KH,PO,

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Vogel's trace elements:

0.001% CuCl₂ 0.001% MnCl₂·4H₂O 0.001% Na₃B₁O₇·10H₂O 0.001% (NH₄)₄Mo₇O₇·7H₂O 0.004% ZnCl2

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For long term storage A. terreus spores were

35 -70°C. suspended in SSS (10%-glycerol, 5% lactose) and stored

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grown on the following solid growth medium (ACM) for 3 to For the generation of spore stocks A. nidulans was 4 days at 37°C.

2% Difco Bacto malt extract 0.1% Difco Bacto peptone 2% glucose 2% agar (Difco, Detroit, MI)

For strains which required para-aminobenzoic acid

(PABA) for growth, PABA was added to a final 10

respectively. Spores were suspended in Tween 80 - saline solution (0.025% Tween 80, 0.8% NaCl) and stored at 4°C. uracil and uridine these were added at 20 mM and 10 mM, concentration of 1 $\mu g/ml$. For strains which required

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0.6\$ (w/v) NaNO, 0.52\$ (w/v) KCI 0.52\$ (w/v) KH.PO, 0.052\$ (w/v) MgSO, 7H,O 1\$ (w/v) glucose 0.1\$ (v/v) ANM trace elements solution PH to 6.5 and make up to 1 liter with distilled

was added. If required the glucose can be omitted and an For preparation of plates 2% (w/v) Difco Bacto agar transformation plates KCl was added at 4.47% (w/v) (0.6 alternative carbon source (e.g., lactose added at the same concentration). For the preparation of . Ξ 52

AMM trace elements solution: 30

.015% (w/v) MnSO4.4H2O 0.1% (w/v) Fe 0.88% (w/v) 2 0.04% (w/v) 0 0.015% (w/v) 0.01% (w/v) N

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0.005% (NH,) MO,O24.7H2O distilled water to 1 liter

Large scale genomic DNA preparation from A. terreus for genomic library construction.

- The mycelium was A 2.5 ml aliquot of spore suspension (10° c.f.u./ml) and rinsed extensively with water then TSE [150 mM NaCl, harvested by filtration through Miracloth (Calbiochem) was used to inoculate 500 ml of liquid CM medium and grown for 20 hours at 30°C and 200 rpm. ß
- 100 mM Na, EDTA, 50 mM Tris-HCl pH 8.0]. The mycelium was chilled pestle and mortar followed by transferral to a liquid nitrogen then ground to a fine powder in a presqueezed dry, broken into small pellets and frozen in 500 ml flask. Fifty ml of extraction buffer [150 mM 10
- NaCl, 100 mM Na, EDTA, 50 mM Tris-HCl pH 8.0, 2% (w/v) SDS) and 10 ml of toluene was added to the flask which was supernatant was removed and extracted with an equal This mixture was centrifuged at 1000 x g for 15 minutes and the shaken at 60 rpm for 72 hours. 15
- This mixture was centrifuged at 10,000 x g for 30 minutes at 15°C. The aqueous layer was carefully removed and 1.1 spooled out from the resulting suspension and resuspended volume of chloroform:isoamyl alcohol (24:1 vol/vol). The DNA was volumes of ethanol was layered on top.

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chloroform:isoamyl alcohol (24:1) and the DNA was spooled in 5 ml TE [10 mM Tris-HCl pH 8.0, 1 mM EDTA] + 50 $\mu g/ml$ out as before. Following resuspension in 1 ml of TE the RNase and 100 µg/ml proteinase K then incubated at 37°C for 2 hours. The mixture was extracted again with 25

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DNA was extracted once with phenol:chloroform:isoamyl

alochol (25.24.1 vol/vol), once with chloroform:jsoal

alcohol (25:24:1, vol/vol), once with chloroform:isoamyl alcohol (24:1) and precipitated with 0.6 volumes isopropanol. The DNA clot was removed, dried briefly and resuspended in 0.5 ml TE.

Small scale genomic DNA preparation from A. terreus for Southern blot.

A 0.5 ml aliquot of spore suspension (10° c.f.u./ml)
was used to inoculate 100 ml of liquid CM and grown for
20 hours at 30°C and 200 rpm. The mycelium was harvested
by filtration through Miracloth (Calbiochem) and rinsed
extensively with water then TSE [150 mM NaCl, 100 mM
Na;EDTA, 50 mM Tris-HCl pH 8.0]. The mycelium was
squeezed dry, broken into small pellets and frozen in
15 liquid nitrogen. The mycelium was ground to a fine
powder in a pre-chilled pestle and mortar and transferred
to a mortar pre-heated to 65°C. Three ml of lysis buffer
[0.5 M NaCl, 10 mM Tris-HCl pH 7.5, 10 mM EDTA, 1% (w/v)
SDS] at 65°C was added and 0.3 ml of 10% (w/v)

20 cetyltrimethylammonium bromide in 0.7 M NaCl. After thorough mixing to form a slurry, 3 ml of phenol:chloroform:isoamyl alcohol (25:24:1) was added. This mixture was transferred to a Corex tube and incubated at 65°C for 15 minutes. Following

25 centrifugation at 12,000 x g for 15 minutes at 4°C the aqueous phase was carefully removed and re-extracted once with phenol, once with phenol:chloroform:isoamyl alcohol (25:24:1) and once with chloroform:isoamyl alcohol (24:1). The DNA was precipitated from the extract by

00037629 PCT/US9979583 addition of 0.1 volume of 3 M sodium acetate pH 5 and 0.6 volumes isopropanol then collected by centrifugation (10,000 x g, 10 minutes, 4°C). After washing with 70% ethanol the pellet was briefly dried and resuspended in TE + RNase (50 μ g/ml).

Transformation of A. terreus.

A 0.5 ml aliquot of spore suspension (10° c.f.u./ml) was used to inoculate 100 ml of liquid CM and grown for 20 hours at 30°C and 200 rpm. The mycelium was harvested by centrifugation at 2000 x g for 15 minutes at 4°C and washed twice with an aqueous solution containing 0.27 M CaCl, and 0.6 M NaCl. To produce protoplasts the washed mycelia was resuspended in 20 ml of the same solution containing 5 mg/ml Novozym 234 (NovoNordisk) and

incubated at 30°C for 1 - 3 hours with gentle agitation.

Protoplasts were separated from undigested mycelia by

filtration through Miracloth (Calbiochem). The

protoplast suspension was diluted with an equal volume of

STC1700 [1.2 M sorbitol, 10 mM Tris-HCl pH 7.5, 35 mM

20 NaCl] and incubated on ice for 10 minutes. The protoplasts were collected by centrifugation (2000 x g, 10 minutes, 4°C), washed with STC1700 and resuspended in 1 ml STC1700. Plasmid DNA, purified using Qiagen columns, (2 - 5 μg in 10 μl) was added to 150 μl of protoplast suspension-and incubated at room temperature

protoplast suspension-and incubated at room temperature for 25 minutes. PEG solution [60% (w/v) polyethylene glycol 4000, 50 mM CaCl₂, 10 mM Tris-HCl pH 7.5] was added to the DNA/protoplasts mixture in three steps: 250 µl,

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250 μ l, and 850 μ l with mixing after each addition. The suspension was incubated at room temperature for 25 minutes then diluted to 10 ml with STC1700. Protoplasts were collected by centrifugation as above and diluted

- were plated onto osmotically stabilized plates (CM medium containing 3% (w/v) Difco Bacto agar and 23.4% (w/v) mannitol, 15 ml of agar per plate). After 4 hours growth at 30°C, 25 ml of OL agar [1% (w/v) Difco Bacto peptone, 1% (w/v) Difco Bacto peptone, 1% (w/v) Difco Bacto peptone,
 - 10 1% (w/v) Difco Bacto agar, 200 μg/ml Zeocin] was overlayered onto each dish. The plates were incubated for 3 4 days at 30°C before transformant colonies were picked. These were streaked to single colonies twice on selective media (CM + 100 μg/ml Zeocin) before spore

suspensions were prepared.

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Fransformation of A. nidulans.

A 0.5 ml aliquot of spore suspension (10° c.f.u./ml) was used to inoculate 100 ml of YEPD [2% (w/v) Difco Bacto yeast extract, 2% (w/v) glucose, 0.1% Difco Bacto peptone] liquid medium including necessary supplements and grown for 20 hours at 37°C and 200 rpm. The mycelia was harvested by centrifugation (2000 x g, 10 minutes, 4°C) and washed twice with 0.6 M KCl. To generate protoplasts the mycelia was resuspended in 20 ml of 0.6 M KCl containing 5 mg/ml Novozym 234 and incubated at 30°C for 1 - 2 hours with gentle shaking. Protoplasts were separated from undigested mycelia by filtration through Miracloth (Calbiochem). The protoplasts were harvested

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by centrifugation as described above and washed twice
with 0.6 M KCl, then resuspended in 10 ml 0.6 M KCl + 50

mM Cacl₂. After counting in a haemocytometer the
protoplasts were harvested by centrifugation as before

- 5 and resuspended to a final concentration of 5 x 10⁴ protoplasts/ml. To 50 μl of protoplast suspension, 5 μl of DNA (2 5 μg, purified using Qiagen columns) was added, then 12.5 μl of PEG solution [25% (w/v) PEG 6000, 50 mM Cacl₂, 10 mM Tris Hcl pH 7.5] and the mixture was
- 10 incubated on ice for 20 minutes. A further 0.5 ml of PEG solution was added and the mixture was incubated on ice for a further 5 minutes. A 1 ml aliquot of 0.6 M KCl + 50 mM CaCl, was added and the protoplasts were plated out in 50 μ l, 200 μ l, and 400 μ l aliquots. For
- 15 transformation to uridine prototrophy, protoplasts were plated out onto AMM + 0.6 M KCl plates without adding uridine or uracil supplements. Plates were incubated at 37°C for 3 4 days when transformants were picked. For transformation to hygromycin B resistance protoplasts
 - incubated for 4 hours at 30°C. 30 ml of 1% peptone, 1% agar, 1 mg/ml hygromycin B was then used to overlay the plates, which were incubated for 3 4 days when
- methods were streaked out to single colonies on selective media (i.e., lacking uridine/uracil supplements or containing 1 µg/ml hygromycin B) twice before spore suspensions were made.

transformants were picked. Transformants from both

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Analysis of strains for lovastatin production.

10 Ç described above. Fermentation Method B involved seed culture was used to inoculate 25 ml of FM in a 250 SEED medium in 250 ml unbaffled flasks and grown for 18 Series 25 Incubator Shaker. 0.5 ml of spore suspension (10° c.f.u./ml) and growing at inoculating 50 ml of RPM in a 250 ml unbaffled flask with ml unbaffled flask and grown for 6 days in the conditions suspension (10° c.f.u./ml) was inoculated into 25 ml of of lovastatin production. In Method A, 0.5 ml of spore 25 incubator/shaker). A 1 ml portion of the resulting hours at 250 rpm and 30°C (New Brunswick Scientific Model and 250 rpm for 7 days in a New Brunswick Scientific Two fermentation methods were used for the analysis

15 SEED medium:

0.5% (w/v) Sigma corn steep liquor 4% (w/v) comato paste 1% (w/v) oat flour 1% (w/v) glucose 1% (v/v) Vogel's trace elements distilled water to 1 l

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4.5% (w/v) glucose
2.4% (w/v) Sigma peptonized milk
0.25% (w/v) Difco Bacto yeast extract
0.25% (w/v) polyethylene glycol 2000
distilled water up to 1 l

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RPM:

S 30 1.3% (w/v) rapeseed meal
1.2% (w/v) KNO,
1.3% (w/v) KN,PO,
1.3% (w/v) MGSO, 7H,O
1.05% (w/v) NaCl
1.05% (w/v) Sigma antifoam B
1.05% (v/v) trace elements solution
1.05% (v/v) trace to 1 1 with distilled water. (w/v) lactose

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Trace elements solution is:

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0.16% (w/v) MnSO₄ 0.34% (w/v) ZnSO₄·7H₂O 0.2% (w/v) CoCl₃·6H₂O 0.5% (w/v) FeSO₄·7H₂O

made up to 1 liter with distilled water.

10 acetate, and shaking the mixture on a New Brunswick the media to 3 with HCl, adding an equal volume of ethyl The cultures were extracted by adjusting the pH o F

20 15 monacolin J, monacolin L, and dihydromonacolin L (acid gun. Extracts were compared with authentic lovastatin, phosphomolybdic acid in methanol and heating with a heat run on C-18 reverse phase TLC plates (RP-18 F254 - Merck) of methanol. For TLC analysis 10 μl of this extract was dried under a nitrogen stream and resuspended in 0.1 ml hours. For analysis, 1 ml of the ethyl acetate layer was Scientific Series 25 incubator/shaker at 250 rpm for 2 (9:1). TLC plates were developed by spraying with 10% in a solvent system of methanol: 0.1% phosphoric acid

30 of acetonitrile (B) and 0.1% phosphoric acid (A). The C_{i*} (3.9 x 150 mm) column was used with a solvent system and lactone forms). For HPLC analysis a Waters Nova-Pak Array Detector; lovastatin was detected at 238 nm. For metabolites were detected with a Waters 996 Photodiode Millenium Software) with a flow rate of 1.5 ml/min and column was eluted with a preprogrammed gradient of 0 to (7.8 x 300 mm) column was used. The same solvent system purification of metabolites a Waters Prep Nova-Pak HR C_{IB} 100% B into A over 25 minutes using gradient 7 (Waters

above was used with gradient of 0 to 100% B in A over

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75 minutes at a flow rate of 4.5 ml/min. Fractions were collected manually, back extracted with ethyl acetate and dried. For HPLC-MS an Aquapore OD-300 7 micron (1.0 x

5 acetonitrile into A (0.05% TFA) over 30 minutes at a flow rate of 0.02 ml/min.

100 mm) column was used with a gradient of 0 to 100%

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CLAIMS

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We claim:

1. A method of increasing the production of lovastatin in a lovastatin-producing organism, comprising the steps of transforming the organism with the D4B segment, wherein the segment is transcribed and

5 translated, and wherein an increase in lovastatin production occurs. 2. The method of claim 1 wherein the D4B segment is the A. terreus D4B segment.

3. The method of claim 1, wherein the D4B segment is identical to nucleotides 579 - 33,000 of SEQ ID NO:18 and 1 - 5,349 of SEO ID NO:19.

4. The method of claim 1, wherein the lovastatin-producing organism is selected from the group consisting of A. terreus ATCC 20542 and ATCC 20541.

5. The method of claim 1, wherein the organism is selected from the group consisting of fungi and yeast.

 The method of claim 1 wherein the increase is at least 2-fold.

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WO 00/37629 PCT/US99/29583 7. The method of claim 1 wherein the nucleic acid

sequence is identical to a sequence isolated from ATCC 98876.

- 8. The method of claim 1 additionally comprising transforming the organism with the entire A. terreus lovastatin gene cluster.
- 9. The method of claim 8 wherein the gene cluster comprises SEQ ID NOS:18 and 19.
- 10. The method of claim 8 wherein the nucleic acid sequence of the gene cluster is identical to sequences isolated from ATCC 98876 and 98877.
- 11. A method of increasing the production of monacolin J in a lovastatin-producing organism, comprising the steps of transforming the organism with the D4B segment, wherein the segment is translated, and wherein an increase monacolin J production occurs.

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12. A method of increasing the production of lovastatin in a lovastatin-producing organism, comprising the step of transforming the organism with the LovE gene, wherein the nucleic acid sequence is translated, and wherein an increase in lovastatin production occurs.

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 13. The method of claim 12 wherein the increase is at least 2.0-fold.
- 14. The method of claim 13 wherein the increase is at least 5-fold.
- 15. The method of claim 12 wherein the nucleotide sequence of the LovE gene comprises SEQ ID NO:27.
- 16. A method of increasing the production of lovastatin in a lovastatin-producing organism comprising the steps of transforming the organism with a nucleic acid sequence comprising a truncated version of the A. terreus D4B segment, wherein the nucleic acid sequence is transcribed and translated and wherein an increase in lovastatin production occurs.
- 17. A method of increasing the production of lovastatin in a lovastatin-producing organism comprising the steps of transforming the organism with a nucleic acid sequence comprising a truncated version of the A.
- 5 terreus lovastatin-producing gene cluster, wherein the nucleic acid sequence is transcribed and translated and wherein an increase in lovastatin production occurs.

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18. A method of increasing or conferring the production of monacolin J in a non-lovastatin-producing organism comprising the steps of transforming the organism with a nucleic acid sequence comprising the D4B segment, wherein the nucleic acid sequence is transcribed

and translated and wherein an increase in monacolin J

production occurs.

- 19. The method of claim 18 wherein the D4B segment is the A. terreus D4B segment.
- 20. The method of claim 18 wherein the D4B segment. comprises nucleotides 579 33,000 of SEQ ID NO:18 and 1-5,349 of SEQ ID NO:19.
- The method of claim 18 additionally comprising the step of converting the monacolin J into lovastatin.
- 12. The method of claim 18 additionally comprising the step of transforming the organism with a nucleic acid sequence comprising the LovF gene, wherein the nucleic acid sequence is transcribed and translated and wherein
 - 5 an increase in lovastatin production occurs.
- 23. An isolated nucleic acid sequence selected from the group consisting of SEQ ID NOs:20 - 36.

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24. A lovastatin-producing organism, wherein the organism has been genetically modified to have increased lovastatin production, wherein the increase is at least 2-fold.

- 25. The organism of claim 24, wherein the organism is a yeast or a fungi.
- 26. A non-lovastatin producing organism, wherein the organism has been genetically modified to produce monacolin J.
- 27. The organism of claim 26, wherein the organism is a yeast or a fungi.
- 28. A non-lovastatin producing organism, wherein the organism has been genetically modified to produce lovastatin.
- The organism of claim 28 wherein the organism is a yeast or a fungi.

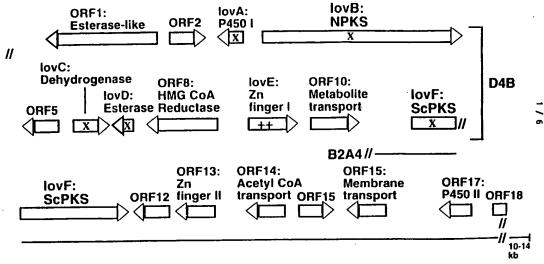
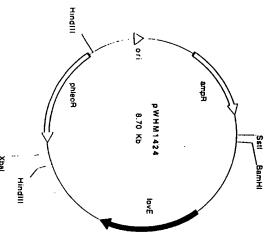


Fig. 1

CB2A4 / PWHM1265 51.00 Kb

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Xbal

ORF10 iovE 6/6 cD48 / pWHM1263 45.50 Kb NPKS / lov8 lovA

FIG. 6

FIG. 5

Wisconsin Alumi Research Foundation Hutchinson, Charles R. Rennedy, Jonathan n.m.: Park, Cheonseck n.m.:

<120> METHOD OF PRODUCING ANTIHYPERCHOLESTEROLEMIC AGENTS

<130> 960296.95718

<160> 36

<170> Patentin Ver. 2.0

<210> 1 <211> 1529 <212> PRT <213> Aspergillus terreus

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PCT/US99/29583 Asp Asn Phe Ala Asp Asn Lys His Pro Asn Asp Tyr Gly Tyr Ser Gln 225 Glu Thr Cys Asp Lys Glu Tyr Gly Ser Gly Val Tyr Ala Gly Gly Phe Thr Gln Gln 220 275 Gly Ser Gly Glu Asp Asp Gly lle Tyr Arg His Asp Ser Glu Tyr Ser 290 Lys Asp Asp Asp Glu Leu His Phe Phe Phe Gly Arg Leu Tyr Thr Arg 335 Ala Tyr Asp Asp Met Met 11e Phe His Lys Asp Lys Asp Ser Gly Ala 345 Val Thr Phe Val Ser Tyr Thr Asn Asn Val His Thr Glu Glu Gln Glu 365 Gly Val His Phe Iie Asp Ile Asn Gly Asp Gly Leu Asp Asp Tyr IIe 385 Phe Thr Lys Gly Gly Thr Phe Ser Thr His Asn Asn Cys Asn Pro Gly 370 380 Cys Ile Ala Leu Asp Gly Thr Thr Tyr Ala Ser Ile Asn Asn Gly Asp 415 Gly Asp Ala Lys Ser Asn Lys Pro Pro Ser Phe Thr Asp 11e Gly Leu 420 Trp Lys Ser Pro Glu Gly Tyr Asp Gln Ala His Val Arg Leu Ala Asp 415 lle Asp Gly Asp Gly Arg Ala Asp Tyr Cys Gly Leu Ala Asp Asn Gly 450 Asp Val Thr Cys Trp Arg Asm Gly Trp lle Glu Asp Ile Pro Ala Tyr 465 Arg Cly Val Arg Phe Clu Asp Ile Asn Gly Asp Gly Arg Asp Asp Trp 500 Phe Ala 575 Lys Val Met Gly Asp Leu Met Trp Val Asp Asp Gly Ala Thr Thr Tyr Thr Asn Ser Arg 515 lle Lys Gly Glu Ser Gly Asp Gly Leu Asn Val Val Trp Arg 535 Gly Phe Tyr Gln Asp Ala Asn Ser Gly Pro Ser His Pro Gly Met 550 Met Ala Asp Ile Trp Tyr Asn Ala Ile Tyr Asn Ala Ala Val Ala 256 Gly Ala Leu Phe Thr Val Arg Ala Gly Lys Gly Ala Ala Asp Pro 305 Leu Ile Val Lys Pro Ala Asp Leu Asp Ile Ser Ser Thr Gly 260 Val lie Phe Gly Thr Ser Gly Leu Arg Asp Gin Val Tyr 570 Trp Gln Pro Leu Gly Lys Arg Phe Thr Gly 490

Tyr Tyr Val.His Val Trp Lys Ser Lys Gly Ala Gly Gly Ala Lys Ile $610\,$

Pro Asn Arg Gly Leu Val Glu Val Pro Ala Asp Gly Ser Ser Phe Trp 660 670 Met Met Asp Tyr Ile Trp Ile His Ser Thr Gly His Met Arg Leu Tyr 645 655 Lys Ala Asp Gly Asp Arg Tyr Cys Asn Met Met Gly His Asp Asn Gly 625 630 Tyr Val Phe Ile Lys Lys Asp Thr Ser Asp Lys Tyr Phe Gly Pro Leu 595

ASH Cys Val Leu Thr Met Trp Trp Tyr Ser Leu Glu Gin Tyr Arg Gin 1265 - 1270 - 1280 Thr Leu Th: Ile Gly Gly Asn Asp Val Phe Phe Ser Asp Leu Val Ser 1250 Lys Ile Asp Gin Trp Leu Gly Gin Asp Fro Thr Gly Thr Thr Met Ala 1235 Tyr The Asn Tyr Ala Cys Ser Gly Asp The The Val Gly Leu Asn Lys 1225 1230 Ser Tyr Gly Lys Leu Val Gln Glu Trp Phe Asp Thr Glu Asp Phe Thr 1215 Met Gly Thr Sly Thr Thr Gly Asp Ser Cys Arg Val Gly Ser 1185 $$1\!\!$ Tyr Gly Val Asn Asp Tyr Val His Phe Gly Asp Ser Tyr Aia Aia Gly 1170 1175 Ser Ser Cys Pro Ala Tyr Asp Asp Ser Ser Tyr Asp Ala Asp Thr Val 1155 Gly Arg Gln Ile Pro Leu Pro Glu Glu Ser Ala Ser Ser Ala Asp Asp 1140 1150 Asp Ser Trp Aia Leu Val Val Leu Gly Gly Tyr Tyr Thr Lys Iie Cys 1135 Gly Asp Ala Asn Trp Gln Arg Arg Leu Leu Cys Pro Asp Pro Asn Asn 1090 Glu Arg Gly Arg Lys Ala Glu Arg Ala Ala Asn Glu Leu Arg Ile Ala 1085 Leu Gly Gln Glu Gly Ile Cys Asp Ser Lys Leu Ser Ala Tyr Asn Ala 1105 - 1110 Lys Gly Aia Arg Asp Leu Aia Ala Gly Thr Phe Asn Arg His Cys Ile 1060 1070 Asp Glu Leu Lys Ser Ser Arg Arg Thr Arg Asp Tyr Val Tyr Gly Trp 1055 Met Met Asn Asn Arg Asn Arg Leu Asn Phe Cys Pro Lys Phe Phe Thr 1025 1030 Ile Thr Cys Lys Asn Leu Arg Gly Cys Asp Glu Asn Gly Trp Leu Ala 1010 1020 Val Thr Met Ile Ser Gly Ser Ser Gln Phe Asp Asp Glu Lys Phe Thr 995 Asp Ser Leu Thr Phe Thr Ile Asn Val Arg Tyr Thr Phe Asp Arg Met 980 980 Gly Ala Tyr Asp Pro Tyr Arg Asp Ile Phe Phe Ala Glu Ser Leu Thr 965 970 Met Gln Tyr Ala Leu 945 Asp Ala Ala Ser Glu Val Ala Tyr Phe Ser Gly 950 955

Asp Asn Ser Giu Cys Thr Arg Ala Gln Val Gln Thr Leu Phe Glu Glu 930 940

Gly Pro Gln Gly Pro Lys Tyr Arg Gly Ala Val Glu Gly Ser Cys Thr $850\,$

Tyr Phe Pro Asp Leu Asn Gly Asp Gly Arg Ala Asp Met His Ser Ile 865 870

Trp Tyr Asn Leu Gly Gln Arg Asp Ile Lys Gly Ser Arg Tyr Glu Trp 835

Thr Lys Asp His Thr Gly Asp Asp Gly Pro Ile Thr Asn Pro Asn Lev 900 905 Trp Asn Ser Ile Asn Asn Thr Ala Gln Thr Trp Tyr Asn Glu Cys Ala 885 890 895

Ser Pro Val Lys Ala Pro Ile Glu Leu Thr Pro His Tyr Gln 915 920 925

Lys Ala Asp Met Ile Trp Thr Asp Lys Phe Ser Gly Asp Gly Ser Val 820 825 Glu Lys Asp Arg Ala Asn Leu His Trp Ala Asp Val Asn Gly Asp Gly 805 816 Asn Gly Asp Asp Gly Trp Asp Tyr Ile Asp Gln Phe Lys Tyr Ser Glu 785 790 800 Ala Asp Tyr Leu Cys Val Glu Lys Asp Gly Arg Thr Trp Gly Trp Val 770 780 Phe Phe Asp Arg Pro Val His Phe Ala Asp Val Ser Gly Asn Gly Lys 755 Ile Asn Ala Ala Asp Glu Leu Tyr Cys Pro Glu His Arg Gly Leu Gly 745 740 Trp Arg Asn Lys Ile Lys Asp Thr Gly Ser Phe Asp Trp Asp Tyr Asn 735 Cys Asp Ile Ile Trp Thr Asp Pro Asp Asn Leu Asn Arg Ala Gln Val705 710Leu Asp Arg Arg Asp Leu His Leu Ala Asp Trp Asp Gly Asp Gly Ala $690\,$ Gly Ala Asn Glu Ile Ile Phe Asp Pro-Gln Glu Gln Ile Gly Met Lys 675 680

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Irp Cys Leu Glu Thr Glu Glu Lys Ala Arg Asn Leu Met Gln Asg Thr Gly Ser Asp Gly Leu Gly Ser Lys leu Arg Ala Ala Tyr Glu Lys Ile 1306 1306 Leu Asp Arg. Ser Gly Ser Ser Val Tyr Leu Pro Val 11e Leu Ile Tyr 1315 Ser Cys Arg Ala Val Leu Arg Arg Ala Asp Phe Thr Leu Val Val Gln Pro Leu Arg Pro Trp Leu Cys His Leu Leu Gln Arg Arg His His Arg Leu Arg Leu Asn His Leu Leu Glu Leu Asn Asp Leu Val Arg Met Leu Asn Ser Leu Ile Gln Ser Thr Ile Ser.Asp Ile Asn Thr Ala Arg Asn Thr Glu Gln lle His Tyr Ile Asp Met Asp Ala Arg Phe Asp Gly His Arg Trp Cys Glu Pro Gly Thr Gln Glu Pro Asp Pro Asp Asn Pro Asn Leu Met Asn Ser Gly Ser Ile Gln Leu Pro Asp Ala Asp Thr Cys Gln Thr Tyr Phe Phe Leu Ser Ala Trp Pro Asp lle Ala lle Val Gly Asp Thr Thr Ala Glu Ser Thr Asn Ala Thr Glu Thr Asp Glu Ile Thr Ala Asp Ala Leu Gly Ser Asp Pro Asp Pro Tyr Ala Val Phe Met Cys Asp Val Ala Val His Val Lys Ala Asn Ser Ser Leu Ile Ala Gln Ser Leu Asp Arg Ala Asn Gln Ala Ile Ala Asn Arg Asp Tyr Ser Ser Gln 1505 Asp Val Ser Trp Trp Leu Pro Ser Pro

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10

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Trp Pro Val Ile Leu Arg Pro Leu Val His Trp Leu Glu Pro Gln Gly

45

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Gly Ser Leu Met Leu Ala Tyr Leu Leu Ile Arg Tyr Asp Trp Lys Val Val Pro Asp 275Pro Arg Ala Cys Pro Gly Arg Phe Phe Ala Ser Lys Glu Ile Lys Met 260 265 270 Ala Gln Leu Glu Asn Thr Asn Gly Asp His Ile Gly Phe Gly Trp His $250\,$ Pro Tyr Arg Tyr Met Arg Leu Arg Glu Asp Pro Ala Lys Ala Phe Ser 235 235 Asp Arg Met Ser Asn Pro Glu Val Trp Pro Glu Pro Ala Lys Tyr Asp 210 215 Ser Asn Gly Thr Phe Ile Pro Lys Gly Glu Leu Val Ala Val Ala Ala 195 200 205 val Glu Cys Ala Thr Met Arg Ser Tyr Ala Leu Gln Asp val Thr Phe 180 185 Leu Lys Leu Leu Asp Ser Cys Leu Lys Glu Ser Gln Arg Val Lys Pro $165\,$ Arg Thr Val Ile Gly Gln Gly Gly Trp Thr Pro Ala Ser Leu Tyr Lys 145 Asp Ile Val Arg His Pro His Leu Leu Glu Pro Leu Arg Asp Glu Ile 130 $130\,$ Ala Lys Gly Lys Trp Tyr Asp Ala Ala Gly Aia Gln Leu Ala Met Asp 100 105 Phe Ala Gly 11e Tyr Gly Thr Ser Asp Leu Le: 11e Gly Gly Leu Val 115

Glu Pro Leu Gln Tyr Tyr Arg His Ser Phe Ser Val Arg Ile His Pro 290 $295 \ 300$

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Thr Phe Giu His Ala Ile Tyr Ala Gln Tyr Gln Gln Giy 405

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PCT/US99/29583

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PCT/US99/29583 lis Met Gln His Ash Arg. Thr Ser Lys Asp Ala Ile Ser Gly Thr Glu Tyr Gly Ala ile Arg Thr Pro Val Tyr Val Val ile Leu Giu His Ala Gly Asp Ile His Phe Val Gin Ile Glu Tyr Lys Asn Thr Tyr Leu Arg Arg Lys Val Pro Thr Leu Ser Cys Asn Leu Gly Arg WO 00/37629

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Gly Thr Leu Asp Glu Cys Leu Arg Thr Lys Asn Leu Phe Thr Ala Val $290\,$ Arg Leu Leu Thr Val Leu Arg Gln Gln Ala Gln Ala Asp Cys His Gln 275 His Met Phe

Glu Gly Ser Arg Ser Gln Ser Pro Ser Arg Asp Asp Thr Ser Ser Ser 345 His Cys Tyr Ile Leu Asn Val Arg Ile Leu Thr Ala Ile Ser Glu Leu $305 \hspace{1.5cm} 310 \hspace{1.5cm} 315$ Leu Leu Ser Gln Ile Arg Arg Thr Gln Asn Ser His Met Ser Pro Leu 325

Ala Ala Pro Lys Gln Gln Gln Pro Leu Leu Ile Gly Ile Met Met Gly 145 150 Met Gly Gly Ser Gly Leu Thr Asn Gly Ala Ile Thr Ile Leu Ser Ala 130 135 140 Thr Ala Arg Ser Ser Thr Met Leu Ile Val Gly Arg Ala Val Aia Siy 115 Phe Leu Ala Phe Leu Gly Leu Phe Glu Ile Gly Ser Val Leu Cys Gly
100 105 110 Gln Pro Leu Ala Gly Lys Leu Tyr Thr Leu Leu Thr Leu Lys Tyr Thr 85 90 95 Asp Val Gly Trp Tyr Gly Ser Aia Tyr Leu Leu Ser Ser Cys Aia Leu 65 70 7:

Pro Ile Gly Glu Leu Phe Ser Tyr Val Asp Pro Leu Thr His Ala $370\,$ Ser Gly His Ser Ser Val Asp Thr Ile Pro Phe Phe Ser Glu Asn Leu 355

Ile Ser Met Ser Gly Glu Pro Gly Glu Asp Ile Ala Arg Thr Gly Ala
420
420
425 Asn Glu Ile Thr Leu Gly Val His Ser Ala Gln Gly Ile Ala Ala Ser 410 405 61 u

Phe Ser Ala Cys Thr Thr Leu His Val Gly Val Gln Leu Leu Arg 385 390

Thr Asn Ser Ala Arg Cys Glu Glu Gln Pro Thr Thr Pro Ala Ala Arg

Ser Ala Gly Ser Arg Gly Arg Thr lle Ala Ala Leu Arg Arg Cys Tyr 465 470 480 Val Leu Phe Met Phe Leu Ser Asp Glu Gly Ala Phe Gln Glu Ala Lys $450\,$

Glu Asp Ile Phe Ser Leu Ala Arg Lys His Lys His Gly Met Leu Arg 485 490

Asp Leu Asn Asn Ile Pro Pro 500

<210> 9 <211> 542 <212> PRT <213> Aspergillus terreus Ile Val Thr Ala Ile Pro His Ile Thr Ala Gln Phe His Ser Leu Gly $50 \hspace{0.5cm} 55 \hspace{0.5cm} 60$ Val Ser Val Thr Leu Val Ala 35 Gln Met Gin Ile Asn His Val Thr Gly Leu Arg Leu Gly Leu 20 25 30 <000> 9

Met Thr Ser His His Gly Glu Thr Glu Lys Pro Gln Ser Asn Thr Ala

10
15 Phe Leu Met Leu Leu Asp Met Ser Ile 45 Val Val

Ser Pro Thr Met Ser Gly Val Tyr Met Leu Pro Gly Ile Gly Gly Gln 345Met Tyr Gln Phe Leu Gly Gly Val Gly Arg Gly Cys Gly Met Gln Thr 415Gly Leu Val Ser Thr Phe Gln Pro Glu Thr Ser Ile Ala Ala Trp Val 385 390 400 Tyr Val Pro Trp Ala Leu Ala Ser''Gly Ile Leu Val Ser Ile Ser Ala 370 $375\,$ The Val Met Ala lie Val Thr Gly Ala lie lie Gly Lys Thr Gly Tyr 355Ile Phe Ser Tyr Tyr Leu Pro Ile Tyr Phe Gln Ala Val Lys Asn Val $325\,$ Gly Gly Ala val Ala Met Ile Pro Ile Ser val Ala Ser Arg Arg Gln 290 300 Ala Ala Gly Val Ser Leu Val Leu Phe Gly Cys Trp Glu Arg His Val 275 280 285 Gly Ser Asp Tyr Ala Trp Asn Ser Ser Val Ile Ile Gly Leu Phe 260 265 270 Val Trp Cys Ser Cys Phe Phe Leu Gly Phe Phe Ser Gly Ala Leu Leu 305 310 320 Leu Phe Ala Gly Phe Ala Ile Met Ile Ser Leu Ala Leu Glu Trp Gly 245 255 Arg Gly Ala Arg Asp Val Leu Thr Gln Leu Asp Phe Leu Gly Phe 235 230 235Leu Pro Ser Thr Ser Asp Ser Thr Thr Asp Gly Thr Asn Pro Lys Arg 210 215 Gly Ala Phe Ala Thr Phe Leu Leu Leu Val Ile Gln Ile Pro Asn Arg 195 200 205 Thr Gln His Ala Ser Trp Arg Trp Cys Phe Tyr Ile Asn Leu Pro Ile 180 Leu Ser Gln Ile Ala Ile Val Cys Gly. Pro Leu Leu Gly Gly Ala Phe 175

5

Asn

Thr Met Ser

Ala Ile Gln Ser Let Arg Ala Gly Glu Ser Asp Met Aia lle Val Ala 195

WO 00/37629

Leu Gly Phe Leu Ser Ser Asp Gly ile Ser Tyr Ser Phe Asp Ser 225Ala Asp Gly Tyr Gly Arg Gly Glu Gly Val Ala Ala Ile Val Leu 250

Gly Ala Asn Leu Leu Leu Asn Pro Asp Val Phe Thr 210 210

Thr Leu Pro Asp Ala Val Arg Asp Gly Asp Pro lle Arg Leu lle Val Arg Glu Thr Ala 11e Asn Gln Asp Gly Arg Thr Pro Ala 11e Ser Thr Pro Ser Gly Glu Ala Gln Glu Cys Leu Ile Gln Asp Cys Tyr Gln Lys 290 300 Ala Gin Leu Asp Pro Lys Gin Thr Ser Tyr Val Giu Ala His Gly Thr Gly Thr Arg Ala Gly Asp Pro Leu Glu Leu Ala Val Ile Ser Ala Ala

Pro Val Val Ala ile Gin Asn Ala Leu Pro Fro Gin Thr Ser Pro Ile 420 430

Gly lie Ser Leu Aia Met Phe Gly Gin Thr Phe Gly Gly Ser Leu Phe 435

Lec Leu Thr Leu Thr Glu Leu Val Phe Ser Asn Gly Leu Asp Ser Gly 450

Arg Gln Tyr Ala Pro Thr Leu Asn Ala Gln Glu Val Thr Ala Ala 465 Ala Thr Gly Phe Arg Gin Vai Val Pro Ala Pro Leu Ile Ser Arg 495

Val

Leu Leu Ala Tyr Ser Lys Gly Val Asp His Ala Phe Tyr Val Ala Val 500 510

Gly Ala Ser Gly Ala Thr Phe Ile Phe Ala Trp Gly Met Gly Arg Leu 515 Ala Trp Arg Gly Trp Arg Met Gln Glu Lys Gly Arg Ser Glu 530 540

<210> 10 <211> 2532 <212> PRT <213> Aspergillus terreus

Gly Met Gly Cys Arg Phe Gly Gly Gly Ala Thr Asp Pro Gln Lys Leu 25 30 30

Trp Lys Leu Leu Glu Glu Gly Gly Ser Ala Trp Ser Lys Ile Pro Pro 35

Ser Arg Phe Asn Val Gly Gly Val Tyr His Pro Asn Gly Gln Arg Val $50 \ \,$

Gly Ser Met His Val Arg Gly Gly His Phe Leu Asp Glu Asp Pro Ala 65 Leu Phe Asp Ala Ser Phe Phe Asn Met Ser Thr Glu Val Ala Ser Cys \$95\$

Met Asp Pro Gln Tyr Arg Leu Ile Leu Glu Val Val Tyr Glu Ala Leu 100

Glu Ala Ala Gly 11e Pro Leu Glu Gln Val Ser Gly Ser Lys Thr Gly 115

Phe Ala Gly Thr Met Tyr His Asp Tyr Gln Gly Ser Phe Gln Arg 130 Pro Glu Ala Leu Pro Arg Tyr Phe 11e Thr Gly Asn Ala Gly Thr 150 Met Leu Ala Asn Arg Val Ser His Phe Tyr Asp Leu Arg Gly Pro Ser $170\,$ Val Ser 11e Asp Thr Ala Cys Ser Thr Thr Leu Thr Ala Leu His Leu 180

Phe Pro Gly Gln Ile Gln Val Gly Ser Val Lys Ale Asn Ile Gly 345 His Thr Glu Ala Val Ser Gly Leu Ala Ser Leu Ile Lys Val Ala Leu 355Ala Val Glu Lys Gly Val lle Pro Prc Asn Ala Arg Phe Leu Gln Pro 370 Ser Lys Lys Leu Leu Lys Asp Thr His Ile Gin Ile Pro Leu Cys Ser 385 Gln Ser Trp Ile Pro Thr Asp Gly Val Arg Arg Ala Ser Ile Asn Asn 410 Pro Phe Ala Glu Thr Ser 11e Cys Pro Pro Asn Gly Tyr Ser Gly Asn 435 Phe Gly Phe Gly Gly Ala Asn Ala His Ala Ile Val Glu Gln Tyr Gly $_{420}^{\circ}$ Tyr Asp Gly Asn Leu Gly Thr Asp Gln Ala His Ile Tyr Val Leu Ser 450 Ala Lys Asp Glu Asn Ser Cys Met Arg Met Val Ser Arg Leu Cys Asp 465 Tyr Ala Thr His Ala. Arg Pro Ala Asp Asp Leu Gln Leu Leu Ala Asn 495 Leu Gly Ser Arg Arg Ser Asn Phe Arg Trp Lys Ala $505\,$ val Cys Thr Ala His Ser Leu Thr Gly Leu Aia Gln Asn Leu Ala Gly 515 lle Ala Tyr Thr 500

18

Met Arg Pro Ser Lys Ser Ala Asp Gln Val Arg Leu Gly Trp 535

Gla

19

Leu Val Thr Asp Val Ala Val Phe Asp Glu Ala Asp Pro Val Gly Gly 1225 1230 Asp Met Leu Arg Ala Gln Ala Lys Met His Ser Gln Ser Pro Ser Ala 1210 1215 Ala Asp Thr Ala Ser Ala Met Pro His Ala Tyr Glu Ser Gln His Ile 1140 1145 Thr Cys Ile Glu Ser Asp Gly Arg Gly Ser Trp Cys Thr Phe Ala Ile 1125 1130 1135 Ser Leu His Arg Val Gly Ile Arg His Gly Pro Phe Phe Arg Asn Ile 1105 1110 1115 Asp Pro Arg Pro Trp Ser Arg Lys Thr Ala Pro Gln Glu Leu Trp Asp 1090 1095 1100 Ala Glu Met Asp Gln Pro Pro Ser Ser Leu Ser Asn Gln Gln Arg Ile 1075 1080 1085 Thr Ala Asp Lys Asn Asp Trp Thr Glu His Cys Thr Gly Leu Val Arg 1060 1065 Gln Ser Leu Gly Ser Gln Asp Trp Gln Arg Phe Leu Val His Ser Ile 1045 1055 Tyr Ile Leu Arg Asp Val Asn Phe Ala Gln Ala Leu Ile Leu Pro Ala 1010 1015 1020 Gly Ile Ser Thr Leu Cys Ser Ser Asp His Glu Ser Asp Asp Ile Ser 995 1000 1005 Val Leu Arg Val Ser Asp Leu Pro Trp Leu Arg Asp His Val Val Gly
975
979 Gly Leu Gln Glu Pro Leu Asn Leu Pro Leu Ala Arg Ser Trp His Asn 945 950 960 Phe Pro Arg Gly Cys Glu Ala Aia Arg Val Gln Val Leu Ser Asp Leu 905 910 Val Gly Cys Met Lys Ile Ser Ser Arg Leu Ala Asp Leu Glu Ala Arg 1185 - 1190 - 1200 Leu Pro Phe Ala Gly Ser Arg Ile Lys Ser Ala Met Val Pro Ala Arg 1170 1175 1180 Val His Pro Thr Thr Leu Asp Ser Ala Val Gln Ala Ala Tyr Thr Thr 1155 1160 1165 Asp Gly Glu Glu Gly Ile Asp Leu Arg Leu Thr Ile Cys Ala Pro Asp 1025 1030 1035 Ser His Ile Val Phe Pro Gly Ala Gly Phe Val Cys Met Ala Val Met 980 985 The Ser Gln Ser Ala Arg Gln Arg Lys Gly Pro Val His Asp Leu Ile 930 940 Pro Pro Tyr Pro Trp Asn His Glu Thr Arg Tyr Trp Lys Glu Pro Arg 915 920 925

Ser Glu Leu Ile Arg Ala Gly Phe Pro Val Asp Leu Asn Ala Ile Asr 895 Cys Leu Ser Arg Gly Lys Ser Ser Leu Se: Thr Leu Arg Leu Leu Ala 865 870 870 Ile Met Gln Leu Pro Glu Leu Ala Thr Cys Asp Ile Pro Tyr Leu Ser $850\,$ Val Ile Glu Ile Gly Pro His Gly Ala Leu Gly Gly Pro Ile Lys Gln 835 \$840Arg Arg Met Cys Leu Asp Glu Asn Asp His Met Pro Lys Val Asp Arg 825 830 His Trp Val Glu Cys Met Leu His Pro Val Glu Phe Glu Ser Ala Phe 805 810 815 Arg Thr Gly Ala Arg Leu His Asp Met Asn Arg Leu Arg Asp Pro Ile 785 795 800 Pro Ser Asp Ala Ala Asn Ala Ser Lys Asp Val IIe Tyr Ala Ser Pro 770 770 Met Thr Asp Ala Phe Arg Ala Gly Leu Thr Glu Leu Phe Gly Ala Asp $755\,$ Ala Ile Ala Lys Leu Glu Glu Leu Leu His Ala Asp Arg Ile Phe Ala 725 730 Tyr Ile Lys Glu Met Gly Ser Thr Trp Ser Ile Ile Glu Glu Leu Ser 580 590 Arg Arg Leu Lys Val Thr Gln Ala Phe His Ser Ser His Met Asn Ser 740 745 750Gly Cys Val Asn Ser Pro Ser Ser Val Thr Val Ser Gly Asp Leu Ser 705 715 720 Ile Tyr Ile Arg Gln Val Pro Leu Gln Ser Glu Glu Cys Leu Val Val $690\,$ His Lys Gly Gly Met Leu Ala Val Gly Leu Ser Arg Ser Glu Val Gly 675Ser Tyr Ile Arg Gly Ala Leu Thr Ala Arg Asp Arg Leu Ala Ser Val 660 670 Ile Glu Met Tyr Pro Val Phe Lys Glu Aia Leu Leu Glu Cys Asp Gly 575 Val Phe Thz Gly Gln Gly Ala Gln Trp Phe Ala Met Gly Arg Glo Leu $545 \ \ \, 550 \ \ \, 555$ Ala Ala Tyr Ala Ile Gly Ala Leu Thr Ala Arg Ser Ala Ile Gly Ile $650\,$ Asn Ile GIn Pro Val Ala Val Thr Ser His Ser Ser Gly Glu Ala Ala 625 630 630 635 Leu Ser Thr Ala Leu Gln Ile Ala Leu Vai Arg Leu Leu Trp Ser Trp 610 620 Arg Pro Glu Thr Glu Ser Arg Val Asp Gln Ala Glu Phe Ser Leu Pro 595 600 605

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Pro Val Met Glu Leu Glu Gly Leu Val Phe Gln Ser Leu Gly Ala Ser 1235 1240 1245

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Leu Gly Thr Ser Asp Arg Asp Ser Thr Asp Pro Gly Asn Thr Cys Ser Ser Trp His Trp Ala Pro Asp ile Ser Leu Val Asn Pro Gly Trp Leu Glu Lys Thr Leu Gly Thr Gly Ile Gln Slu His Glu Ile Ser Leu Ile Leu Glu Leu Arg Arg Cys Ser Val His Phe Ile Gln Glu Ala Met Glu Ser Leu Ser Val Gly Asp Val Glu Arg Leu Ser Gly His Leu Ala Lys Phe Tyr Ala Trp Met Gin Lys Gin Leu Ala Cys Ala Gin Asn Gly Glu Leu Gly Pro Glu Ser Ser Trp Thr Arg Asp Ser Glu Gln Ala Arg Cys Ser Leu Arg Ser Arg Val Val Ala Gly Ser Thr Asn Gly Glu Met ile Cys Arg Leu Gly Ser Val Leu Pro Ala Ile Leu Arg Arg Glu Val Asp Pro Leu Glu Val Met Met Asp Gly His Leu Leu Ser Arg Tyr Tyr Val Asp Ala Leu Lys Trp Ser Arg Ser Asn Ala Gin Ala Ser Glu Leu Val Arg Leu Cys Cys His Lys Asn Pro Arg Ala Arg Ile Leu Glu Ile Gly Gly Gly Thr Gly Gly Cys Thr Gln Leu Val Val Asp Ser Leu Gly 1455 Pro Asn Pro Pro Val Gly Arg Tyr Asp Phe Thr Asp Val Ser Ala Gly Phe Phe Glu Ala Ala Arg Lys Arg Phe Ala Gly Trp Gln Asn Val Met Asp Phe Arg Lys Leu Asp 11e Glu Asp Asp Pro Glu Ala Gln Gly Phe Val Cys Gly Ser Tyr Asp Val Val Leu Ala Cys Gln Val Leu His Ala thr Ser Asn Met Gln Arg Thr Leu Thr Asn Val Arg Lys Leu Leu Lys Pro Gly Gly Lys Leu lle Leu Val Glu Thr Thr Arg Asp Glu Leu Asp Leu Phe Phe Thr Phe Gly Leu Leu Pro Gly Trp Trp Leu Ser Glu Glu 1555 Pro Glu Arg Gln Ser Thr Pro Ser Leu Ser Pro Thr Met Trp Arg Ser Met Leu His Thr Thr Gly Phe Asn Gly Val Glu Val Glu Ala Arg Asp

Cys Asp Ser His Glu Phe Tyr Met Ile Ser Thr Met Met Ser Thr Ala /al Gln Aia Thr Pro Met Ser Cys Ser Val Lys Leu Pro Glu Val Leu Leu Val Tyr Val Asp Ser Ser Thr Pro Met Ser Trp Ile Ser Asp Leu Gln Gly Glu 11e Arg Gly Arg Asn Cys Ser Val Thr Ser Leu Gln Ala Leu Arg Gin Val Pro Pro Thr Glu Gly Gin Ile Cys Val Phe Leu Gly Glu Val Glu His Ser Met Leu Gly Ser Val Thr Asn Asp Asp Phe Thr Leu Leu Thr Ser Met Leu Gln Leu Alæ Gly Gly Thr Leu Trp Val Thr $1700\ 1710$ Gln Gly Ala Thr Met Lys Ser Asp Asp Pro Leu Lys Ala Leu His Leu GJy Leu Leu Arg Thr Met Arg Asn Glu Ser His GJy Lys Arg Phe Val $1730\,$ Ser Leu Asp Leu Asp Pro Ser Arg Asn Pro Trp Thr Gly Asp Ser Arg Asp Ala Ile Val Ser Val Leu Asp Leu Ile Ser Met Ser Asp Glu Lys Glu Phe Asp Tyr Ala Glu Arg Asp Gly Val Ile His Val Pro Arg Ala Phe Ser Asp Ser Ile Asn Gly Gly Glu Glu Asp Gly Tyr Ala Leu Glu Pro Phe Gln Asp Ser Gln His Leu Leu Arg Leu Asp lle Gln Thr Pro GJy Leu Leu Asp Ser Leu His Phe Thr Lys Arg Asn Val Asp Thr Tyr Glu Pro Asp Lys Leu Pro Asp Asp Trp Val Glu lle Glu Pro Arg Ala Phe Gly Leu Asn Phe Arg Asp lle Met Val Ala Met Gly Gln Leu Glu Ser Asn Val Met Gly Phe Glu Cys Ala Gly Val Val Thr Ser Leu Ser Glu Thr Ala Arg Thr Ile Ala Pro Gly Leu Ala Val Gly Asp Arg Val 1890 1890 ys Ala Leu Met Asn Gly His Trp Ala Ser Arg Val Thr Thr Ser Arg 1915 thr Asn Val Val Arg lle Pro Glu Thr Leu Ser Phe Pro His Ala Ala 1936 1935 Ser lle Pro Leu Ala Phe Thr Thr Ala Tyr lle Ser Leu Tyr Thr Val

Ala Arg Ile Leu Pro Gly Glu Thr Vai Leu Ile His Ala Gly Ala Gly 1955 1960 1965 PCT/US99/29583

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Val Asp Phe Phe Val Met Leu Ser Ser Leu Val Gly Val Met Gly Gly 2290 2300 Arg Pro Lys Val Gln Gly Ser Trp Asn Leu His Arg Ile Ala Ser Asp 2275 2280 2285 Asp Ala Leu Val Ser Gln Met Thr Ala Asp Gly Phe His Ala Ala Leu 2260 2270 Glu Met Pro Pro Ile Arg Gly Val lie Gln Gly Ala Met Val Leu Lys 2245 2250 2255 Ser Leu Gln Glu Arg Gly Cys Thr Val Ser Val Gln Ala Cys Asp Val 2210 2215 2220 Tyr Leu Ile Ile Leu Ser Arg Thr Ala Azg Val Asp Pro Val Val Thr 2195 2200 2205 Gly Ile Gly Arg Arg Ile Cys Glu Trp Leu Val Asp Arg Gly Ala Arg 2180 2185 2190 Thr Val Ala Pro Asp Asp Ala Val Leu Val Arg Gln Glu Arg Met Pro 2145 2150 2155 Ala Phe Arg Thr Met Gln Ser Gly Gln His Val Gly Lys Ile Val Val 2130 . 2140 Gly Leu Ile His Pro Ile Ser Glu Tyr Pro Met Ser Ala Leu Glu Lys 2115 2120 2125 Gly val Gly Gln Ala Aia Ile Ile Leu Ala Gln Leu Thr Gly Ala Glu 1970 1975 1980 Ala Asp Glu Ser Gln Leu Glu Ala Ala Leu Gln Gln Cys Arg Ala Glu 2225 2230 2230 Leu Phe Leu Lys Pro Asn Val Ser Tyr Leu Val Ala Gly Gly Leu Gly
2165 2170 2175 Phe Gin Ala Met Ser Glu Val Ile Leu Leu Trp Glu Arg Thr Ala Ile 2100 2105 2110 Ser Ser Val Asp lle Leu Tyr Trp Gln Gln Ala Lys Pro Ala Glu lle 2085 2090 Gln Asn Ser Arg Leu Asp Met Ser Thr Phe Val Arg Asn Val Ser Phe 2065 2070 2075 Leu Ala Arg Phe Gly Arg Phe Val Glu Ile Gly Lys Lys Asp Leu Glu 2050 2055 Val Leu Asn Ser Leu Ala Gly Pro Leu Leu Gln Lys Ser Phe Asp Cys 2035 2040 2045 Lys Phe His Leu Asp Pro Asp His Val Phe Ser Ser Arg Asp Ser Ser Ser 2015 Val Phe Thr Thr Ala Gly Ser Glu Thr Lys Arg Asn Leu Leu Ile Asp 1985 1990 2000 Phe Val Asp Gly Ile Lys Thr Arg Thr Arg Gly Lys Gly Val Asp Val 2020 2025

> Gly Ile Lys Tyr Arg Asp Pro Leu Arg Asp Asn His Gly Ala Leu Ser 2420 2425 Gly Pro His Trp Ala His Ala Asp Trp Met Gln Glu Ala Arg Phe Ala 2415 Arg Tyr Lys Ala 2530 Leu Met Glu Gly Arg Thr Ile Ala Lys Val Ala Glu Val Val Leu Gln 2515 2520 2525 Arg Asn Trp lle Thr Ala Lys Phe Asn Val Asp Ile Ser Val Phe Glu 2500 2510 Gln Thr Leu Ala Gly Ile Gly Val Asp Ser Leu Val Ala Ile Glu Leu 2495 2495 Leu Thr Pro Ala Glu Asp Asp Asn Leu His Ala Arg Leu Asn Arg Ala 2435 2440 2445 Ala Pro Thr Arg Pro Ala Val Ile Val Thr Gly Ile Asn Thr Arg Pro 2385 2390 2395 val Leu Asp Val Leu Glu Gln Ala Ile Ser Pro Val Cys Ser Pro Ala 2370 2375 Ala Glu Arg Leu Gln Arg Ile Gly Tyr Gln Pro Leu His Glu Glu Glu 2355 2360 2365 Gly Met Val Gln Ser Ile Gly Tyr Val Ala Glu Thr Asp Ser kia Val 2345 2350 Ala Glu His Arg Met Ala His Asn Gln Pro Ala Val Thr Ile Asp Leu 2325 2330 2335 Ala Gly Gln Ala Asn Tyr Ala Ala Ala Gly Ala Phe Gln Asp Ala Leu 2305 2310 2320 Lys Leu Ile Ser Met Phe Gly Leu Thr Asp Ser Glu Met Ser Ala Thr 2465 2470 2480 Ile Ser Gln Gln Glu Ser Ile Ala Val Ile Met Glu Ala Met Ser Cys 2450 2455 2460

<210> 11 <211> 249 <212> PRT <213> Aspergillus terreus

Pro Lys Ser Leu Pro Ala Ala His Ser Ala Val Ala Ser Cys Leu Thr 65 70 80 His His Leu Arg His Leu Thr Asn Ile Gly Leu Asp Thr Pro Pro Cys Trp Met Lys Arg Gly Tyr Ser Cys Asn Ser Val Arg Thr Asp Asp Lys 35 Ala Glu Aia Ile Arg Tyr Arg Val Lys Thr Gly Val Ser Met Asp Gly
25 30 <000> 11 Met Ala Thr Gin Glu Phe Leu Ser Asp Val Ser Ser Gly Phe Leu Ser 10 15

PCT/US99/29583 Phe Val Frc Prc Asp Fro Cys Glu Asn Trp Glu Ala Leu Gln Val Ala 90 95 Val Ser Leu Leu Phe Ser Phe Tyr Ser Leu Trp Leu Gln Arg Gly Gly 115 ITP Asp Lys Ala Cys Cys Arg Asn Pro Thr Pro Leu Phe Phe 11e Cys Cys Gly Arg Tyr Gly Gly Leu His Arg Val Ser Lys Val Phe Pro Lys 130 Val Trp Prc Asp Asp Met Asp Ser Gln Leu Pro Ser Arg Leu Gln Thr 145 Leu Val Ser Lys Arg Lys Pro Glu Pro Ala Pro Asn Asn Ser Thr Tyr 170 ile Ser Lys Gly Tyr Ala Thr Phe Phe Asn Gln Phe Ser Leu Pro Ser 180 Val Asp Val Thr Gin lie Leu Asn Gin Thr Leu Gin His His Asp Val 200 Glu Thr Ile Asn Leu Asp Cys Gly Ser Gly Leu Leu Thr Leu Arg Thr 210 \$210Gin Leu Arg ile Leu leu lle Gly Lys Pro Lys ile ile Lys Pro Phe 225 Ser Gly Leu Arg Thr Ser Ile Asn Glu 245

<400> 12
Met Glu Ser Ala Glu Leu Ser Ser Lys Arg Gln Ala Phe Pro Ala Cys
15 Asp Glu Cys Arg Ile Arg Lys Val Arg Cys Ser Lys Glu Gly Pro Lys $20 \ \ \, 25$ Cys Ser His Cys Leu Arg Tyr Asn Leu Pro Cys Glu Phe Ser Asn Lys 35 45 Val Ala Arg Asp Val Glu Lys Leu Gly Ser Arg Val Gly Asp Ile Glu 50 60 arg Asp Leu Ser Arg Pro Gln Ser Gln Glu Ser Gly Tyr Thr Ser Ser 95 Thr Ser Ser Glu Glu Cys Glu Val Asn Leu Tyr Ser Gly Lys His Thr $100 \ 100$ His Ala Leu Gin Arg Cys Leu Ser Phe Ile Asp Ala His Gin Gly Phe 65Ser Pro Thr Glu Glu Asp Gly Phe Trp Pro Leu His Gly Tyr Gly Ser 115 Val Ser Leu Val Met Glu Ala Gln Ala Ala Asn Ala Asn Leu Thr 130 140 <210> 12 <211> 742 <212> PRT <213> Aspergillus terreus Phe

Ser Trp Leu Pro Val Asp Met Thr Ser Gly Gin Val Ala Glu Met Val 145 gra Leu Ser Ala Ser Giu Asn Asp Thr Phe Leu Pro Ser Leu Pro Pro Arg Ala Leu 195 Val Glu Pro Ser 11e Asn Glu Tyr Phe Lys Lys Leu His Pro Arg Leu 210Pro Ile Phe Ser Arg Gln Thr Ile Met Asp Ala Val Glu Ser Gln Tyr 225 Thr lle Arg Thr Gly Pro Pro Asp Leu Val Trp lle Thr Ser Phe Asn 250 Cys lle Val Leu Gln Ala Leu Thr Gln Thr Ser Ile Ala Asn Lys Val 265 270 Val Gly Cys Thr Gly Gln Asp lle Pro lle Asp Tyr Met Ile Ile Ser 275Leu Leu Arg Asn Ile Arg Gln Cys Tyr Asn Arg Leu Glu Fhr Leu Val $290\,$ Ala Met Glu Tyr Phe Asp Phe Ala Ile Phe Leu Thr Ile Fhe Ala Gln 335 Lys Pro Arg Leu Ser Asn Ile Arg Ala Leu Phe Cys Leu Ala Leu Val 305 Val Cys Glu Leu Ser Arg Leu Ile Gly Leu His Leu Thr Thr Thr 345 346 Pro Pro Thr Glu Asp Gly Ala Val Gly Asp Gln Pro Lys Asp Leu Phe 365 Trp Ser lle Phe Leu Val Asp Lys His Val Ser lle lle Gly Gly Lys $370\,$ Asp Ser Ala Ala Pro Leu Pro Asn Ala Phe Ala Ala Arg Ileu 415 Ala Cys Leu Leu Pro Ser Tyr Asp Cys Ser Val Pro Leu Pro Pro Tyr 385 Ala Phe lle Leu Glu Glu Ile Tyr Leu Gly Leu Tyr Ser Ala Lys Ser 420 Ser Lys Met Glu Gln Ser Arg Val Arg Arg Arg Ile Arg Ale Ala 415 Asp Pro Asn Arg Pro Leu Glu Glu Tyr Ile Cys Ala Thr Gln Leu 470 Arg Phe Ala Leu Ser Ser Cys Trp Val Leu Leu His Lys Arg Ile Trp 490 Arg Lys Leu Ser Gln Trp His Val Gln His Glu His Val Leu Arg Thr 450 460 Ala Phe Asp Arg Gin Ala Val Ser Ala Val Arg Ser Lys Val 170 Ala Asn Glu Thr Leu Gln Gln Ile Ile Glu Asp Ile Pro Thr 180

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Tyr Vai His Ile Val Glu Glu Asp Gln Pro Ile His Ser Gln Asp 545 550 Phe Asp Ser Ile Val Leu Asn Tyr Ser Leu Ile Ser Phe Met Gly Ile $530\,$ Met Leu Phe Lys Gln Leu Cys Asp Gly Cys Lys Ser Gly Phe Ser Asn 515 520 Ser Gin Glu Arg Gly Ala Val Cys Leu Gln His Ala Arg Asp Cys Leu 500 510

Cys Ser Asp Ile Ala Leu Leu Leu Gln Asn Leu Arg Glu Arg Arg Phe $595 \\ 600 \\ 605$ Arg Ser Ser Ala Ser Ile Ser Tyr Lys Leu Ser Gln Val Ala Ser Arg 585 590 Glu Ile Leu Thr Phe Phe Ala Ile Tyr Thr Asn Arg Ser Ala Ser Asn 565

Thr Tyr Met Asp Tyr 625 Asp Val Ala Asn Ala Ser Thr Ser Thr Thr Ser 630

Thr Gly Ser Ser Tyr Asn Leu Asn Ile . 645

Ser Pro Leu Gly Val Pro Gly 650

Asp Gly Thr Ile Ala Thr Pro Ser Glu Asp Ala Thr Gln Asp Leu Leu 675 Asp Gly Gln Val Trp Asp Ile Tyr Phe Asn Pro Arg Glu Ile Pro Met 660 665 670

Ser Asn Asp Ala Gly Gln Cys Leu Gly Phe Pro Asp Phe Ser Leu Gly 690 695 700

Ser Glu Phe Gly Leu Iie Met Glu Glu Asp Iie Iie Arg Tyr Glu Arg 735 Ile Asp Asn Phe Ser Asp Phe Pro Leu Gly Ile Asp Met Thr Ser Gln 705 710

Leu Leu Asp Arg Pro Val 740

<210> 13 <211> 301 <212> PRT <213> Aspergillus terreus

Glu Ser Ala Lys Thr Arg Ala Gln Leu Lys Arg Arg Asn His Asp Val 50 55 60 Ala Gly Ala Cys Ala Gly Ala Val Glu Ile Ser Ile Thr Tyr Pro Phe 35 40 45 Thr Gln Lys Ala Arg Gly Lys Arg Thr Lys Gly Ile Pro Ala 20 25 <400> 13
401 Ser Lys Val Gln Thr Asn Val Pro Leu Pro Lys Ala Pro Leu Fig. 10 Leu Val

> Val Tyr Glu Lys Val Tyr Lys Phe Leu Thr Gln Pro Asn 290 295 300 Trp Phe Arg Thr Gly Arg Leu Ser Leu Thr Ser Ala Ile Met Phe Pro 275 Lys Thr Leu Leu Arg Asn Glu Gly Ile Gly Val Phe Trp Ser Gly Val 260 265 270 Gin Ser Leu Gin Ala Arg Gin Leu Tyr Gly Asn Thr Phe Asn Cys Val 245 250 Gly Ile Leu Arg Asp Arg Gly Pro Leu Gly Phe Phe Ser Ala Val Gly 175 $175\,$ Arg Lys Val Gly Asn Ala Glu Leu Ser Th: Thr Phe Gly Ala Ile Ala 145 150 150 Gly Ala Ser Val Leu Ala Gly Phe Gly Ala Gly Val Thr Glu Ala Val 115 120 125 Thr Leu Val Gly Thr Thr Leu Lys Ala Ser Val Gln Phe Ala 95 Ala Ala ile Lys Pro Gly Ile Arg Gly Trp Tyr Ala Gly Tyr Gly Ala 65 70 Cys Cys Ala Trp Ser Thr Gln Pro Leu Asp Val Ile Lys Thr Arg Met 225 230 230 Asp Val His Pro Leu Ala Ser Thr Leu Val Gly Ser Val Thr Gly Val 210 215 220 Tyr Asn Glu Leu Ile Gly Leu Ala Arg Lys Tyr Ser Lys Asn Gly Glu 195 Pro Thr 11e Leu Arg Gln Ser Ser Asn Ala Ala Val Lys Phe Thr Val 180 Leu Ala Val Thr Pro Ala Glu Ala Ile Lys Thr Lys Ile Ile Asp Ala 130 $\,$ Asn lie Tyr Arg Ser Ala Leu Ser Gly Pro Asn Gly Glu Leu Ser Thr 100 105 Ser Phe PCT/US99/29583

<210> 14 <211> 490 <212> PRT <213> Aspergillus terreus

Thr Met Ser Phe Glu Pro Pro Gly Ala Cys Arg Val Ile Gly Tyr Gly 65 Ala Trp Val Gly Ala Arg Val Pro Trp Ser Glu Lys Tyr Val Gln Ala 50 55 60 400> 14
Wet Thr Lys Gln Ser Ala Asp Ser Asn Ala Lys Ser Gly Val
10 Asp Val Leu Glu Arg Ala Lys Tyr Leu Ile Leu Asp Gly Ile Ala Cys 35 40 45 Glu Ile Cys His Trp Ala Ser Asn i.eu Ala Thr Asp Asp Ile 20 25 30 15 15 Pro Pro Ala

PCT/US99/29583 Gin Lys Leu Gly Pro Val Ala Ala Ala Met Thr Asn Ser Ala Phe İle $90\,$ Gin Ala Thr Giu Leu Asp Asp Tyr His Ser Giu Ala Pro Leu His Ser 100 110

Ala Ser Ile. Val Leu Pro Ala Val Phe Ala Ala Ser Glu Val Leu Ala 115 $$\rm 125$

Glu Gin Gly Lys Thr 11e Ser Gly 11e Ala Val 11e Leu Ala Ala 11e 130 Val Gly Phe Glu Ser Gly Pro Arg Ile Gly Lys Ala Ile Tyr Gly Ser 145

Asp Leu Leu Asn Asn Gly Trp His Cys Gly Ala Val Tyr Gly Ala Pro 170 Ala Gly Ala Leu Ala Thr Gly Lys Leu Gly Leu Thr Pro Asp Ser 190

Met Glu Asp Ala Leu Gly Ile Ala Cys Thr Gln Ala Cys Gly Leu Met 195

Ala Arg Asn Gly Leu Leu Gly Gly Leu Leu Ala His Gly Gly Tyr Glu 225Ser Ala Gln Tyr Gly Gly Met Val Lys Arg Val Gln His Gly Phe Ala 210

Ala Met Lys Gly Val Leu Glu Arg Ser Tyr Gly Gly Phe Leu Lys Met 255Phe Thr Lys Gly Asn Gly Arg Glu Pro Pro Tyr Lys Glu Glu Glu Val265

Val Ala Gly Leu Gly Ser Phe Trp His Thr Phe Thr Ile Arg Ile Lys 275 Leu Tyr Ala Cys Cys Gly Leu Val His Gly Pro Val Glu Ala Ile Glu 290

Asn Leu Gln Arg Arg Tyr Pro Glu Leu Leu Asn Arg Ala Asn Leu Ser 305

Cys Gly Trp lie Pro Glu Glu Arg Pro lie Ser Ser lie Ala Gly Gin 345 Asn Ile Arg His Val His Val Gln Leu Ser Thr Ala Ser Asn Ser His 335

Met Ser Val Ala Tyr Ile Leu Ala Val Gin Leu Val Asp Gin Gin Cys 360 Leu Ala Gin Phe Ser Giu Phe Asp Asp Asn Leu Giu Arg Pro Giu 370 ren

Val Trp Asp Leu Ala Arg Lys Val Thr Pro Ser His Ser Glu Glu Phe 385 395 400 Asp Gin Asp Gly Asn Cys Leu Ser Ala Gly Arg Val Arg Ile Glu Phe 415 Asn Asp Gly Ser Ser Val Thr Glu Thr Val Glu Lys Pro Leu Gly Val 425

WO 00/37629

PCT/US99/29583 Lys Glu Pro Met Pro Asn Glu Arg Ile Leu His Lys Tyr Arg Thr Leu 435

Leu Ala Gly Ser Val Thr Asp Glu Thr Arg Val Lys Glu 11e Glu Asp 450 Val Leu Ser Leu Asp Arg Leu Thr Asp Ile Ser Pro Leu Leu Glu 475

Leu Asn Cys Pro Val Lys Ser Pro Leu Val 455

<210> 15 <211> 488 <212> PRT <213> Aspergillus terreus

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Gly Ser Gly Gln Asn Glu Pro Glu Lys Lys Gly Arg Asp Ile Pro Leu 20 30 rrp Arg Lys Cys Val Ile Thr Phe Val Val Ser Trp Met Thr Leu Val 35

Val Thr Phe Ser Ser Thr Cys Leu Leu Pro Ala Ala Pro Glu Ile Ala 50 60

Asn Glu Phe Asp Met Thr Val Glu Thr lle Asn Ile Ser Asn Ala Gly 65 Val Leu Val Ala Met Gly Tyr Ser Ser Leu Ile Trp Gly Pro Met Asn 85 90

Lys Leu Val Gly Arg Arg Thr Ser Tyr Asn Leu Ala Ile Ser Met Leu 100

Cys Ala Cys Ser Ala Gly Thr Ala Ala Ala Ile Asn Glu Lys Met Phe 115 ile Ala Phe Arg Val Leu Ser Gly Leu Thr Gly Thr Ser Phe Met Val 130 $140\,$

Ser Gly Gln Thr Val Leu Ala Asp Ile Phe Glu Pro Val Tyr Arg Gly 145

Thr Ala Val Gly Phe Phe Met Ala Gly Thr Leu Ser Gly Pro Ala Ile 165 Gly Pro Cys Val Gly Gly Val 11e Val Thr Phe Thr Ser Trp Arg Val 180

lle Phe Trp Leu Gln Leu Gly Met Ser Gly Leu Gly Leu Val Leu Ser 195 195Leu Leu Phe Phe Pro Lys Ile Glu Gly Thr Ser Glu Lys Val Ser Thr $210\,$ Phe Lys Pro Thr Thr Leu Val Ser lie lie Ser Lys Phe Ser Pro $240\,$ Thr Asp Val Leu Lys Gln Trp Val Tyr Pro Asm Val Phe Leu Ala Val 250

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Ser Ala Trp Glu Ile Cys Pro Leu His Leu Leu Glu Thr Lys Cys Ser 260 265 270 PCT/US99/29583

Cys Arg Lys Gln Lys Asp Leu Cys Cys Gly Leu Leu Ala Ile Thr 275 280 Tyr Ser Ile Leu Thr Ser Ala Arg Ala Ile Phe Asn Ser Arg Phe His 290 295 Gln

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Pro Ile Ile Ala Ala Phe Phe Ala Gly Trp Gly Leu Met Gly Ser Phe 385 390 390 Leu Ile Tyr Gly Trp Thr Leu Gln Glu Asp Lys Gly Gly Met Val Val $370\,$

Ser Ala Val ile Ala Gly Lys Tyr Met Ile Gln Tyr Ser Phe Ser Ala 420 425 Asn Cys Leu Asn Thr Tyr Val Ala Val Glu \not Lla Leu Pro Arg Asn 405 416

Gly Ser Ser Ala Leu Val Val Pro Val Ile Asp Ala Leu Gly Val Gly 435 Thr Ala Ala Ile Ala Arg Trp Gly Ile Asn Met Gln Arg Trp Ala Glu $_{475}$ Trp Thr Phe Thr Leu Cys Val Val Ala Ser Thr Ile Ala Gly Leu Ile 450 \$450

Arg Ala Phe Asn Leu Pro Thr Gln 485

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Val Phe Gin Ala Arg Leu Gly Asn Arg Arg Val Ile Phe Ala Asn Thr 65 70 75 80 Val Lys His Ala Thr Val Ala Arg Lys Trp Ser Lys Glu Phe Gly Pro Pro Glu Ile Fro Gly Val Pro Ile Phe Gly Asn Leu Ile Gln Leu Gly $35 \hspace{1cm} 40 \hspace{1cm} 45$ Leu Thr Arg Tyr Phe Asn Arg Thr Asp lie Pro Lys Ile Lys Gly Ile 25 30 Phe

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